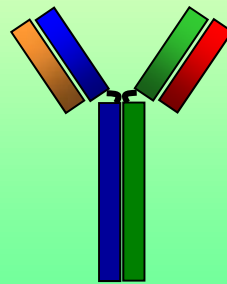
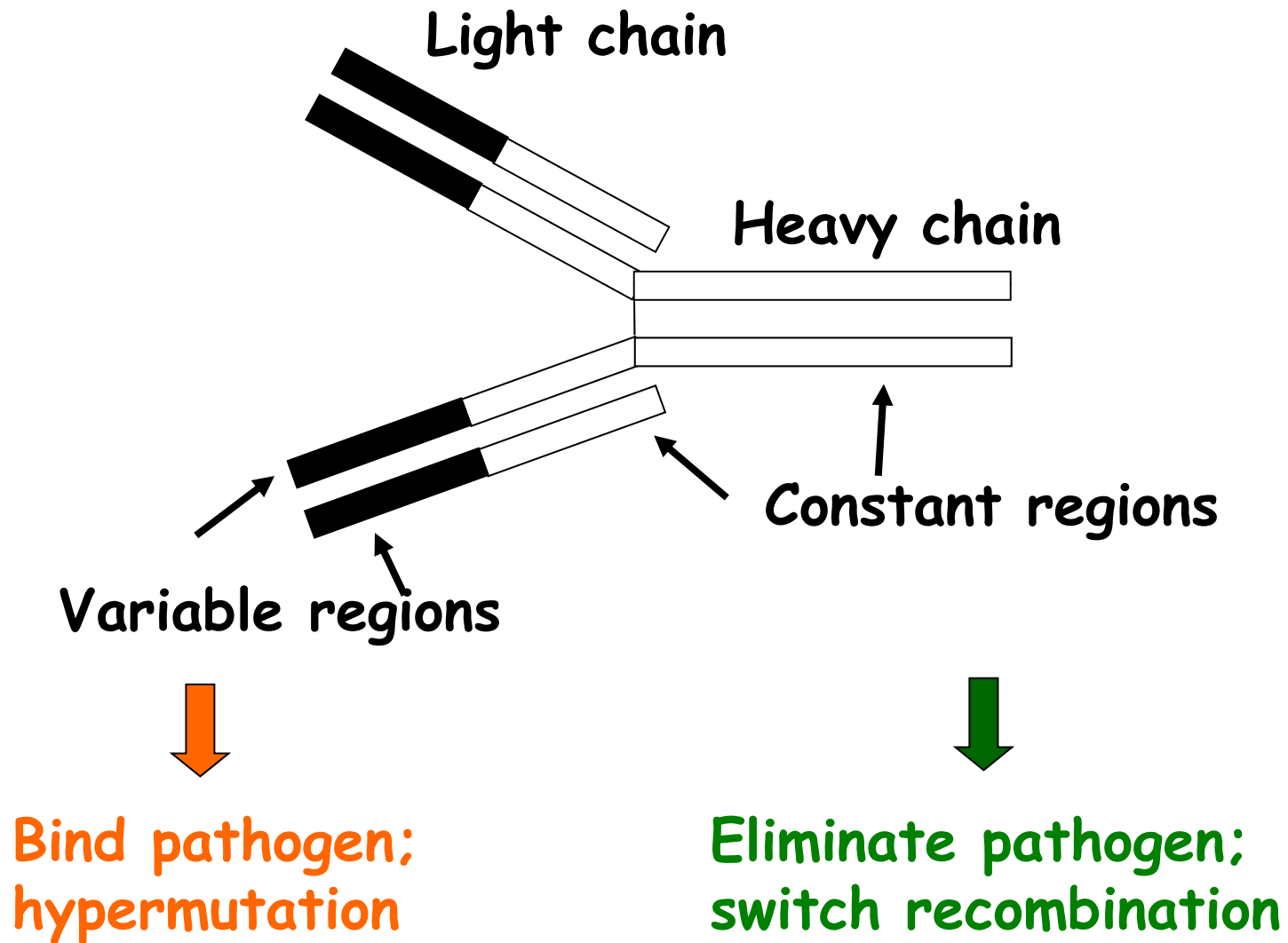


Balancing Somatic Hypermutation and DNA Repair in Immunoglobulin Genes

Patricia Gearhart
Laboratory of Molecular Gerontology
National Institute on Aging, NIH
Baltimore

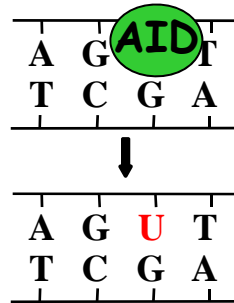


ANTIBODY (B cells)

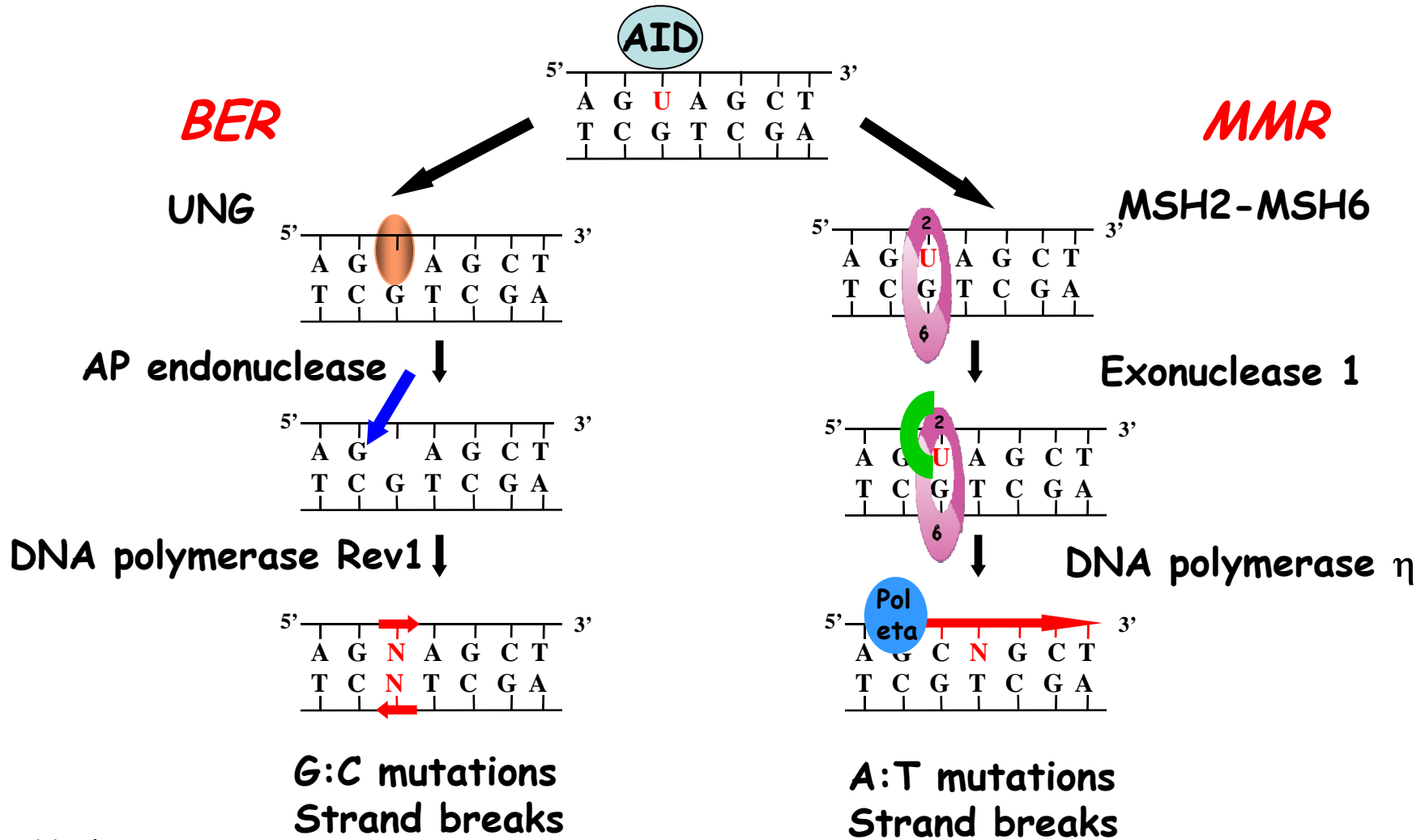


Activation-Induced Deaminase (AID)

deaminates cytosine to uracil



U:G is mutagenic: strand breaks & mutations



Neuberger
Stavnezer
Jacobs

Gearhart
Scharff
Reynaud

I. The Master Catalyst

II. Going Rogue

III. The Targeting Enigma

I. AID - the Master Catalyst

Protein loop that recognizes the AID hot spot:
WRC (W = A/T; R = A/G)

Rahul Kohli, JHU



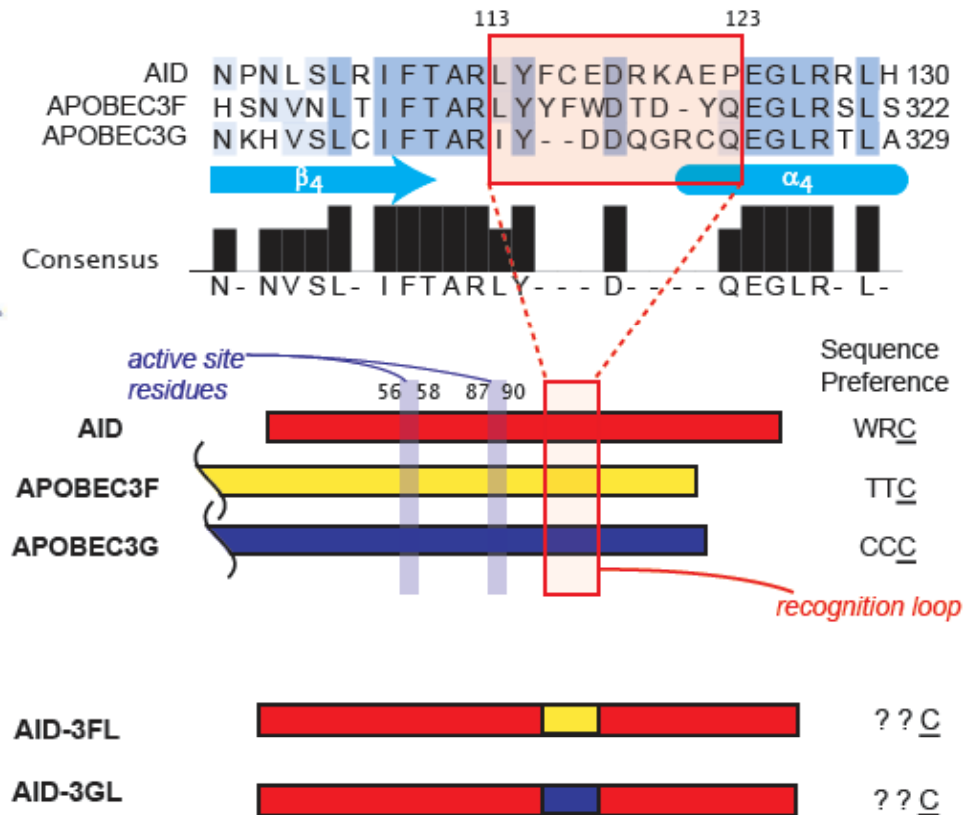
JBC 284:22898-04 (2009)

DNA cytosine deaminases

Antibody diversity:	AID	WRC <u>C</u>
HIV retroviruses:	APOBEC 3F	TT <u>C</u>
	APOBEC 3G	CC <u>C</u>

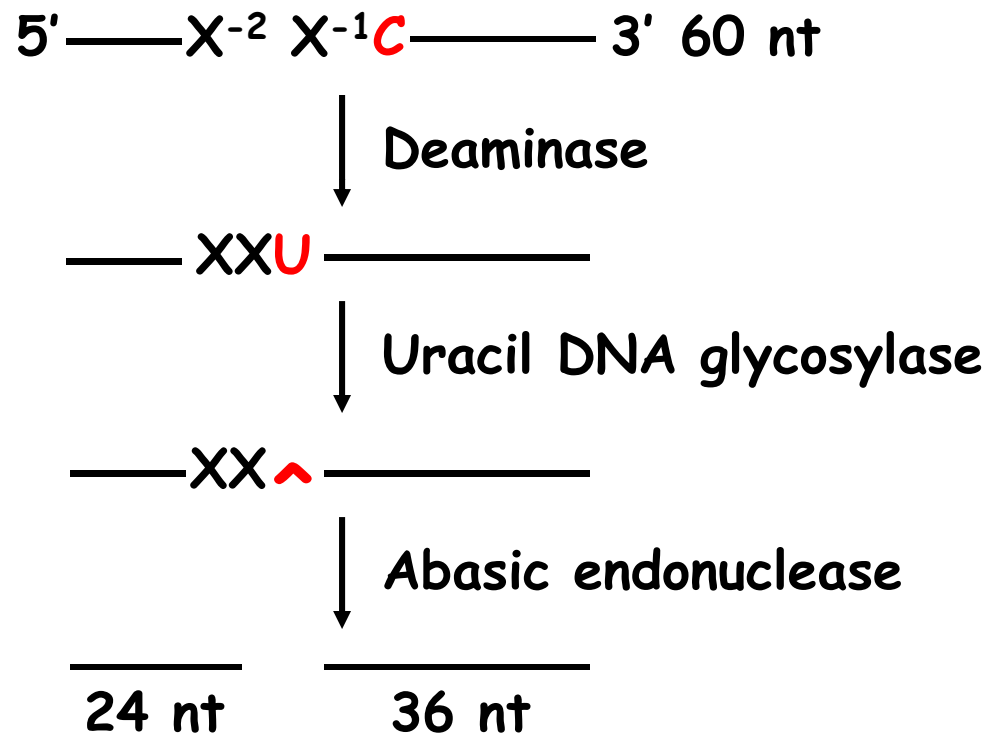
How do they recognize different motifs?

Structural alignment suggests a "hotspot recognition loop"



Hypothesis: Grafting loops from the APOBEC3 enzymes into AID will lead to predictable changes in specificity

Purify variant deaminases from bacterial cultures. Incubate with oligonucleotides containing **XXC** (X = A, mC, G, or T).



AID/APOBEC loop graft variants change
sequence motifs as predicted

AID

WRC

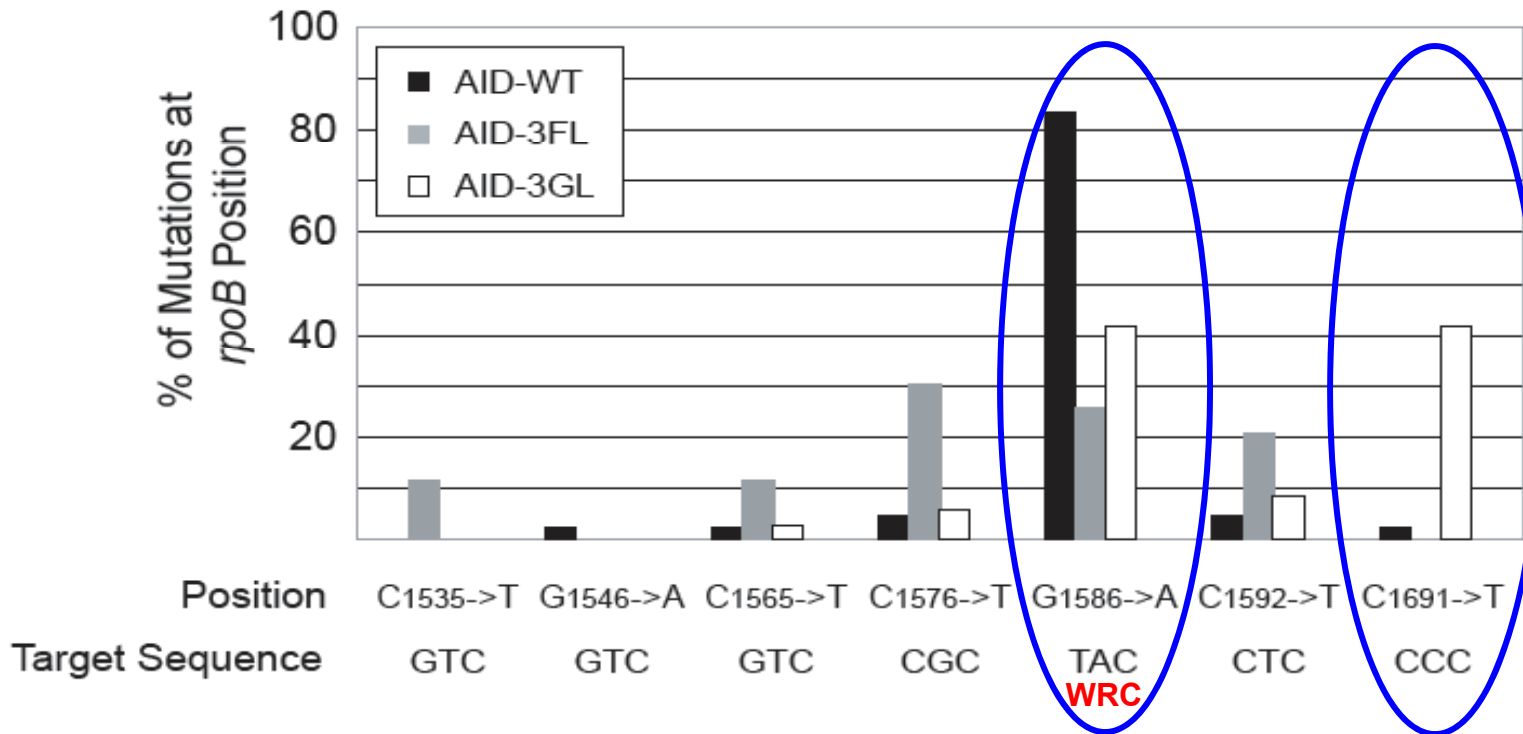
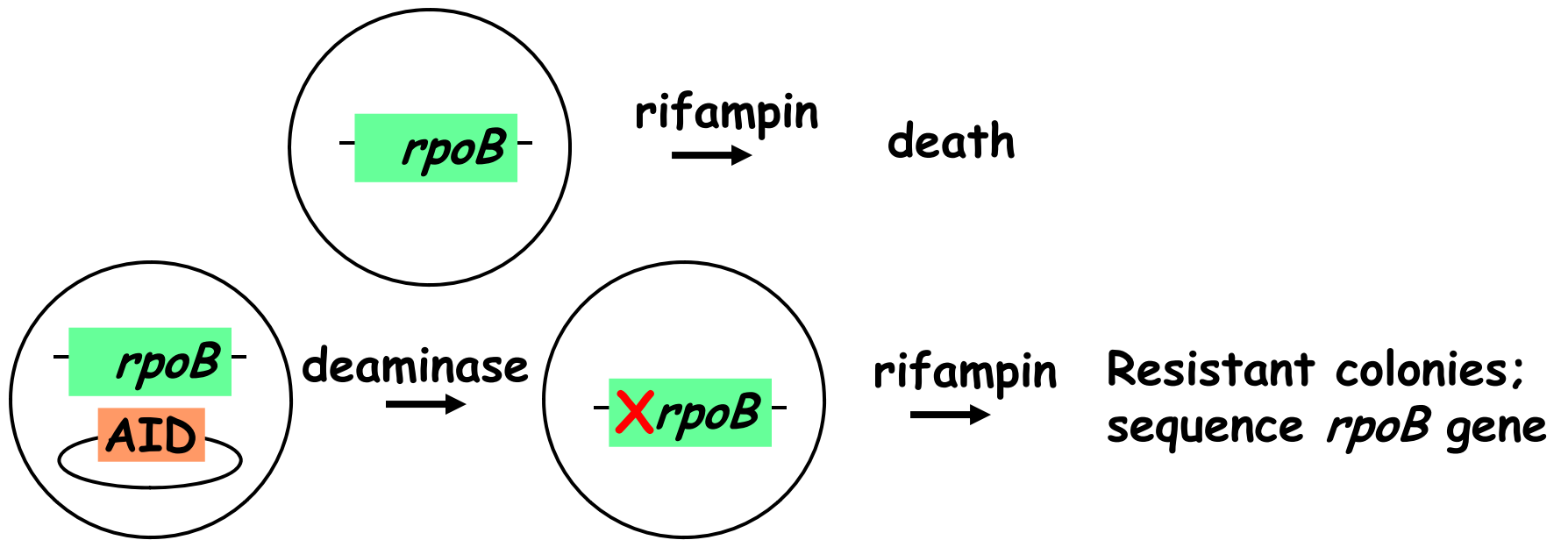
AID-3FL

TTC

AID-3GL

CCC

Are motifs altered in *E. coli*?



9 amino acid loop in AID is responsible for recognizing hot spot motif WRC.

Similar loops in APOBEC3F/G can be transferred to change specificity of AID.

II. Going Rogue - Uracils in DNA

AID proposed to deaminate RNA, whereas genetic and biochemical data support DNA deamination

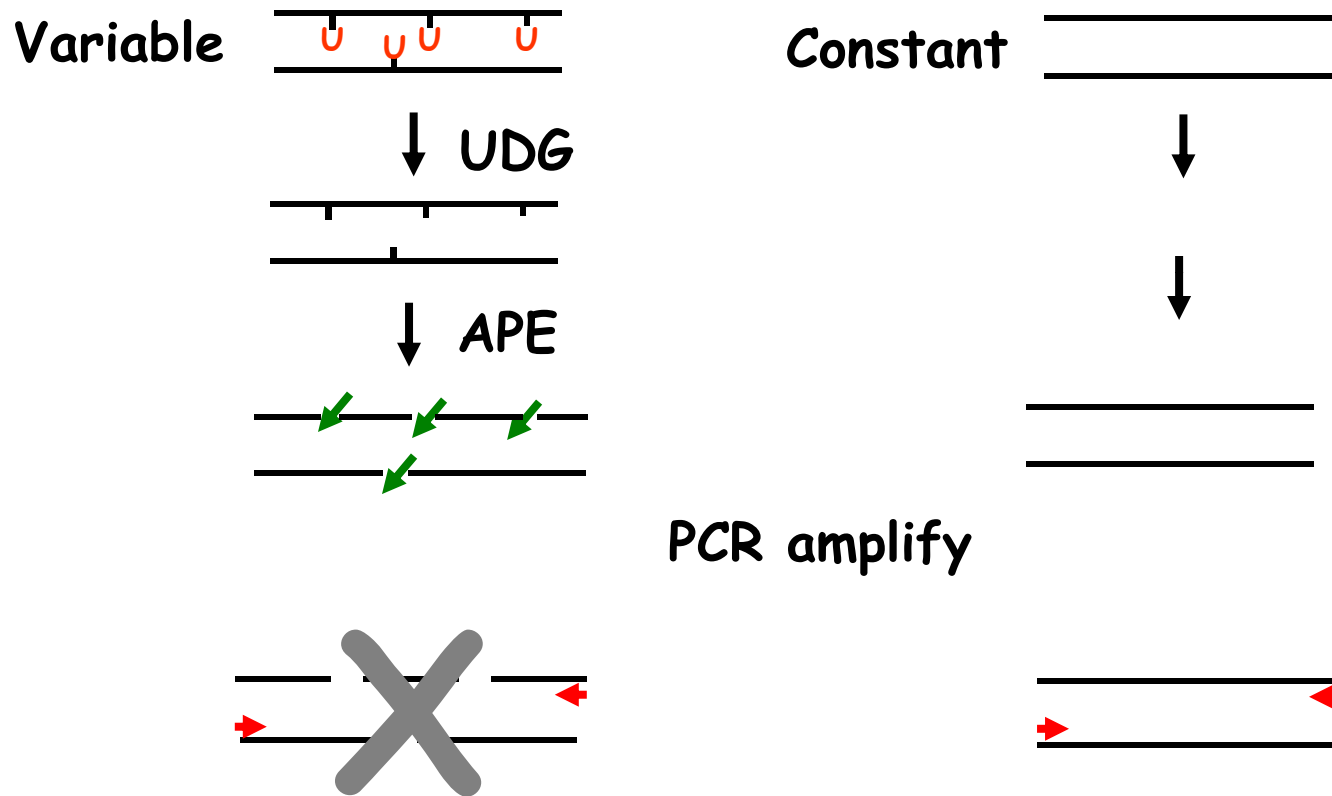
...but, no evidence for uracils in immunoglobulin loci.

Uracils in variable region DNA from B cells

Rob Maul

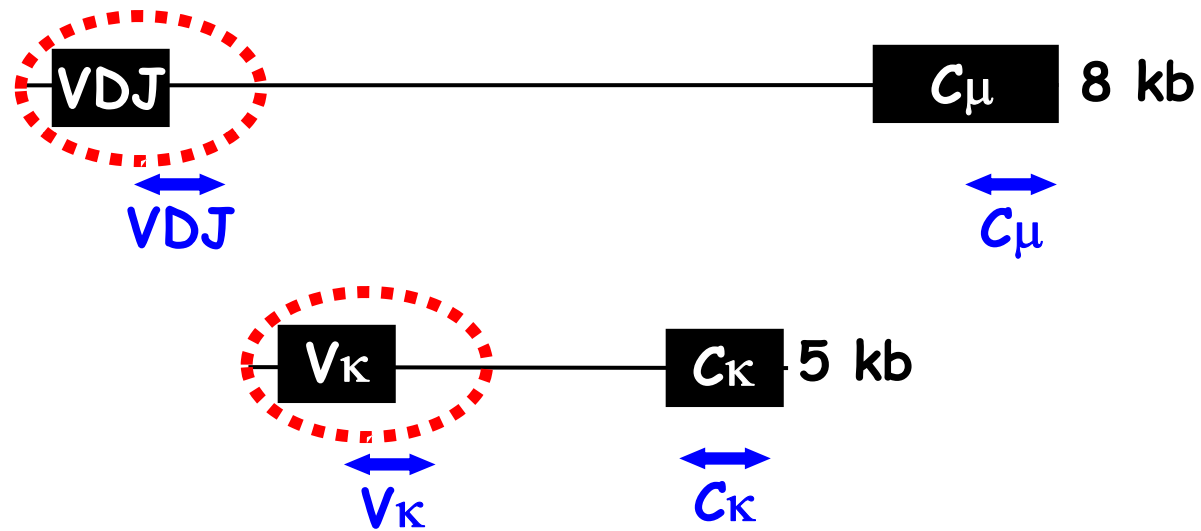


Digest DNA (*Ung*^{-/-} Peyer's patch B cells) with uracil glycosylase (UDG) and abasic endonuclease (APE)

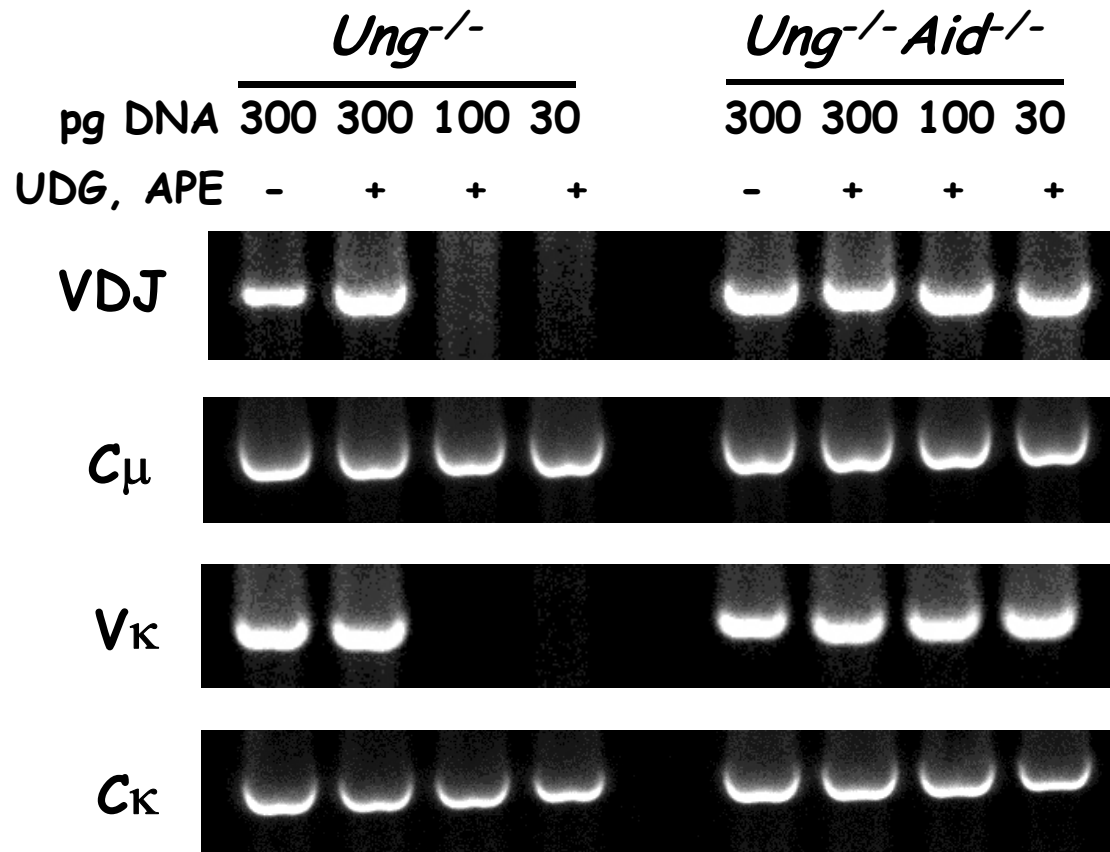


Hypothesis: more uracils = less amplification of V gene

Location of primers for nested PCR



PCR assay



More UDG-sensitive sites in V genes than C genes;
AID dependent.

III. The Targeting Enigma

Setting up the S_{μ} locus for AID

Rob Maul

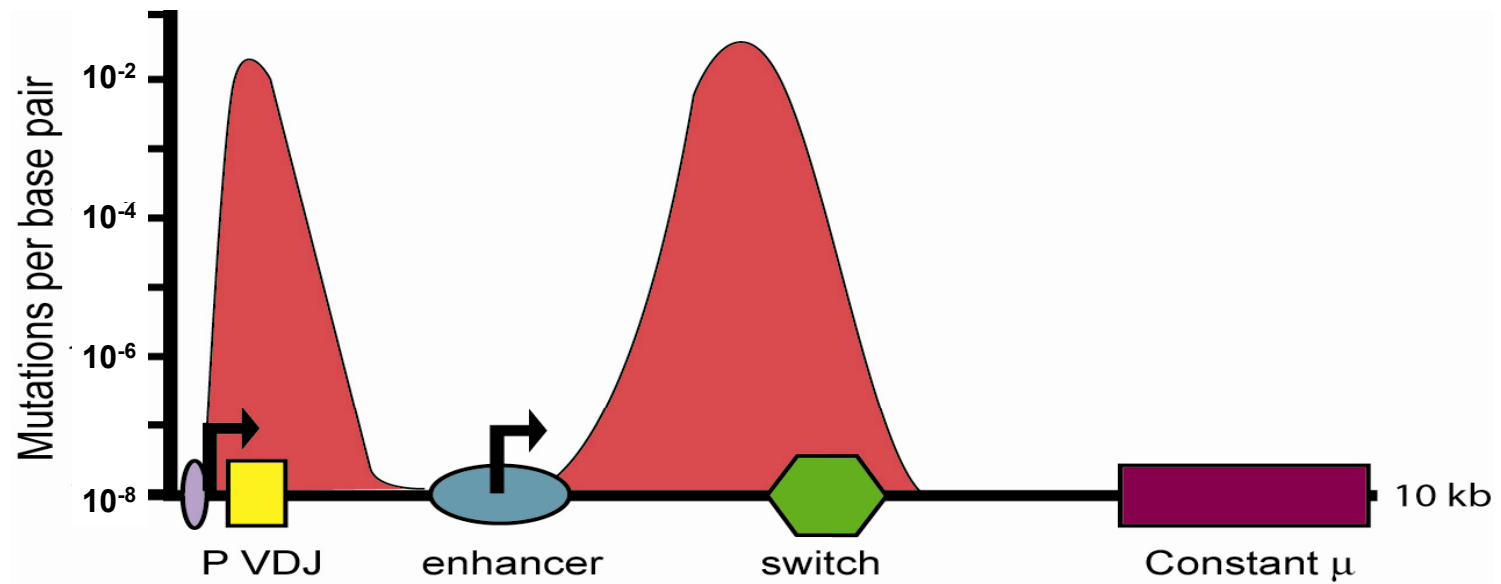


Deepa Rajagopal



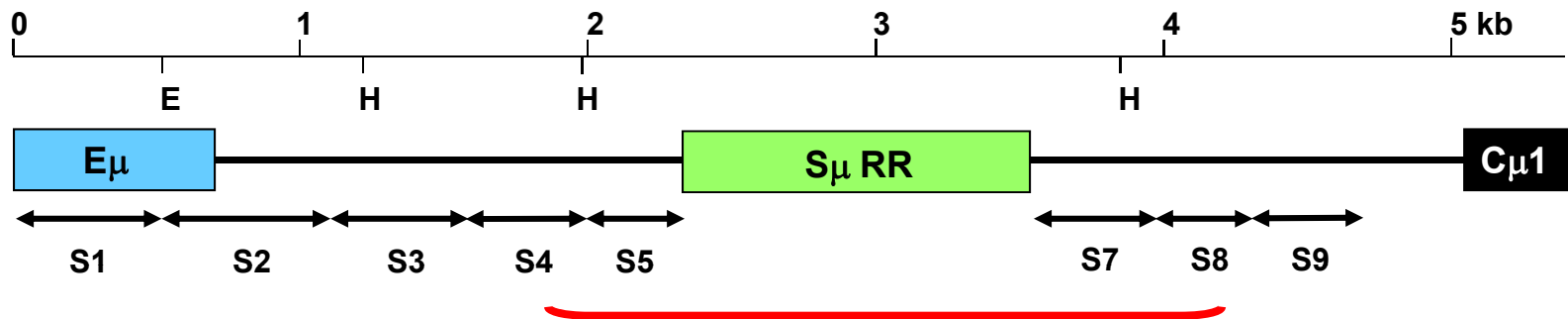
JEM 206:1237-44 (2009)

Peaks of hypermutation after V and I μ promoters



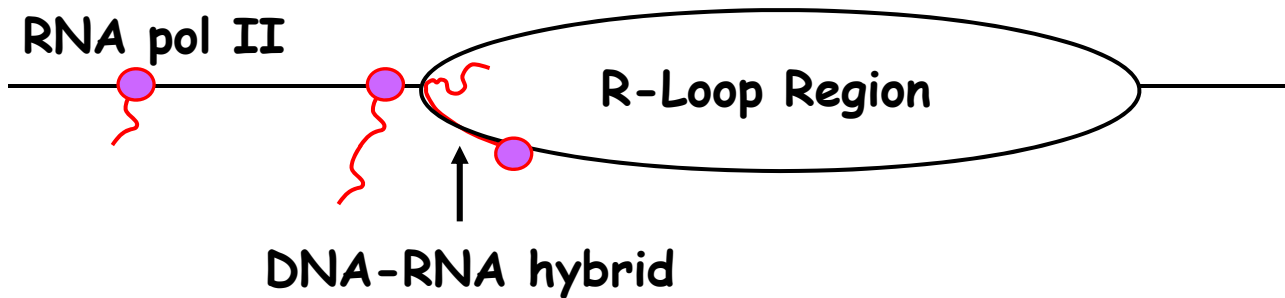
Does location of RNA pol II correlate with this pattern?

How does DNA structure in S_{μ} affect transcription?



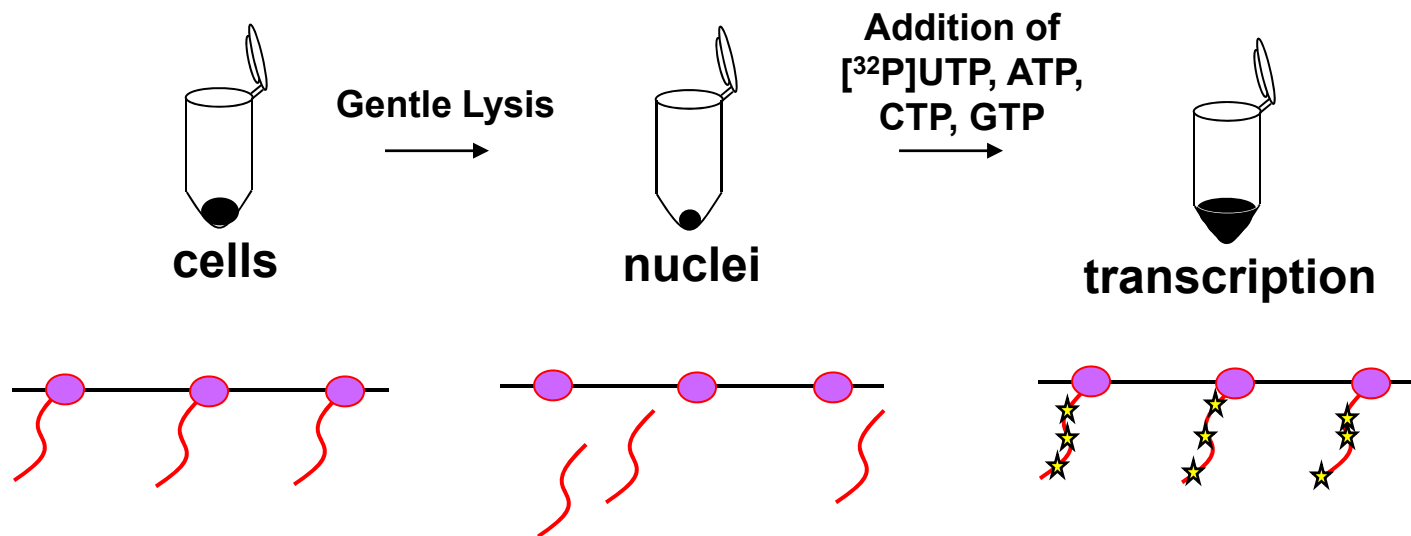
GGGGAGCTGGGGAGCTGGG

Clusters of **G** and **AGCT** hotspots for AID

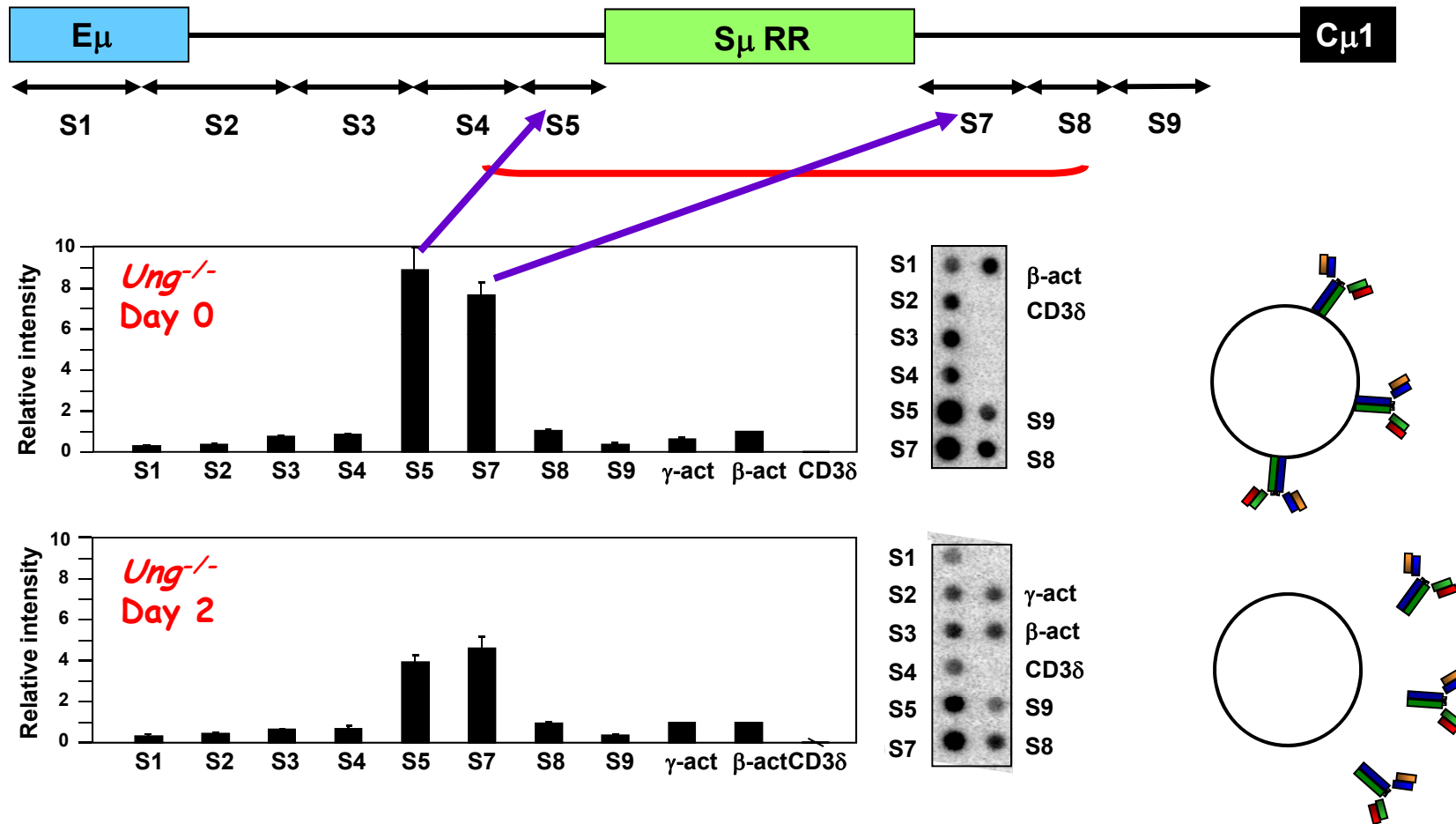


Map the position of RNA pol II by nuclear run-on

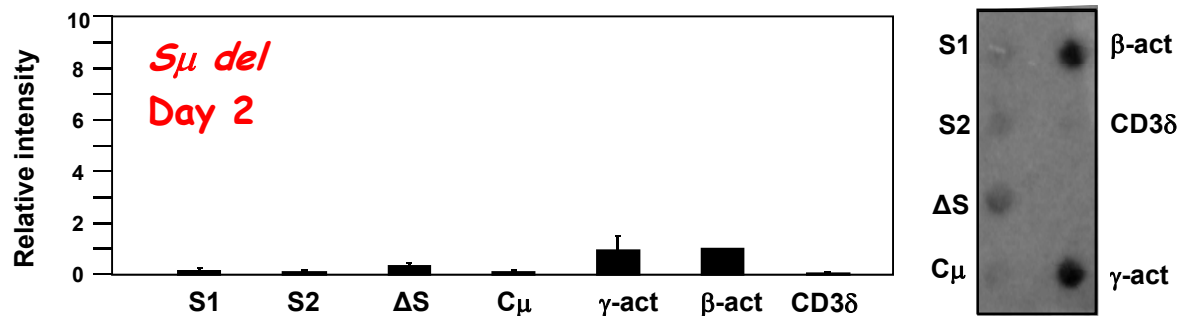
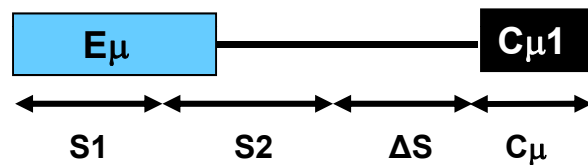
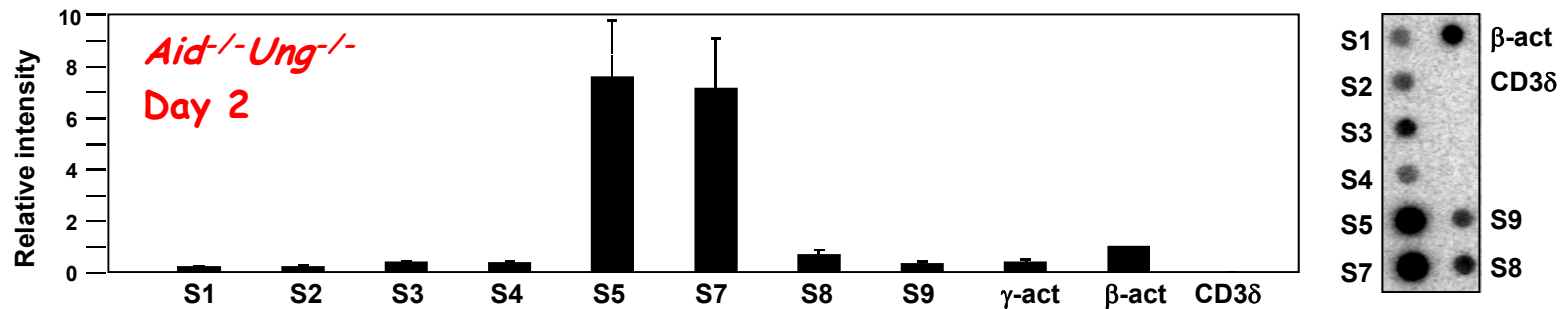
Stimulate *Ung*^{-/-} splenic B cells with LPS + IL4. After 2 days, isolate nuclei.



Position of RNA pol II molecules



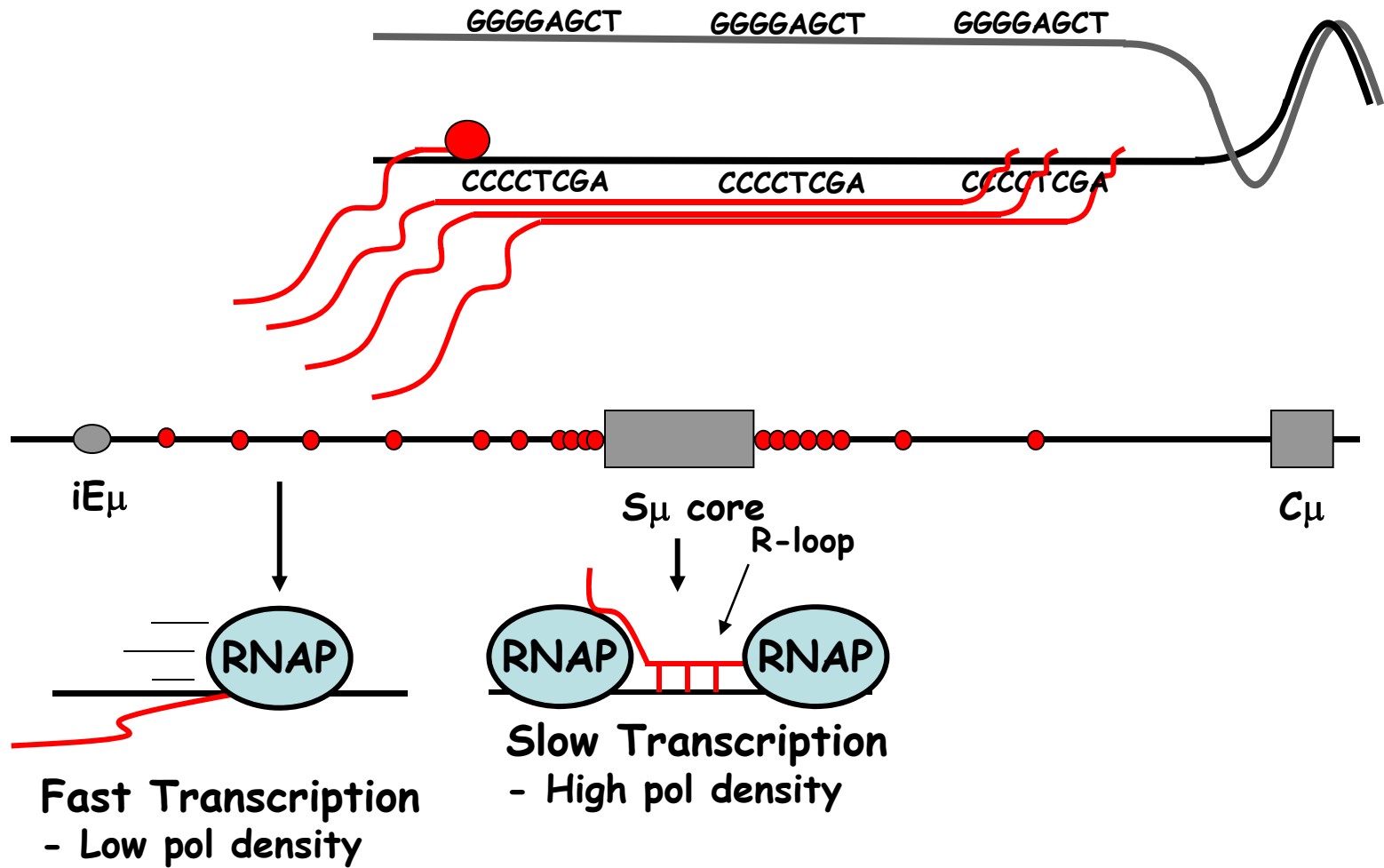
Pileup of polymerases on either side of the repetitive region.
Does not depend on cell activation.



Pileup does not depend on AID. Depends on DNA sequence of S_{μ} .

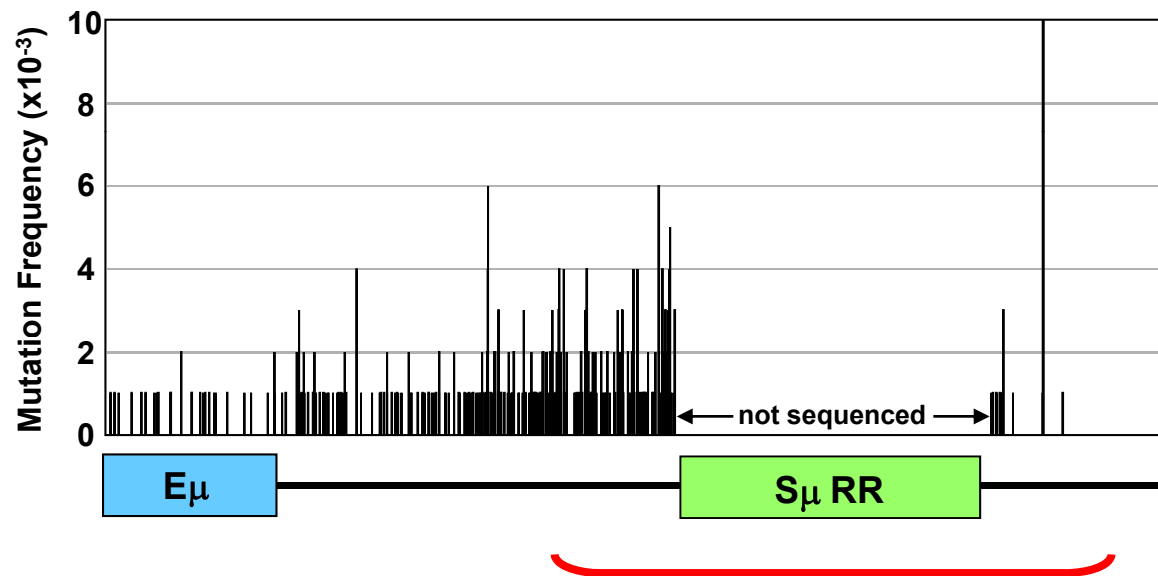
RNA pol II location confirmed by ChIP

RNA pols pause in R-loops



How does DNA structure affect somatic hypermutation?

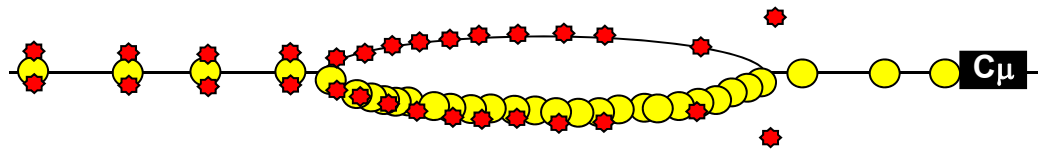
Sequenced J_H4 intron DNA from immunized B cells from $Ung^{-/-}$ mice



Mutations increase before the repetitive region and decrease after

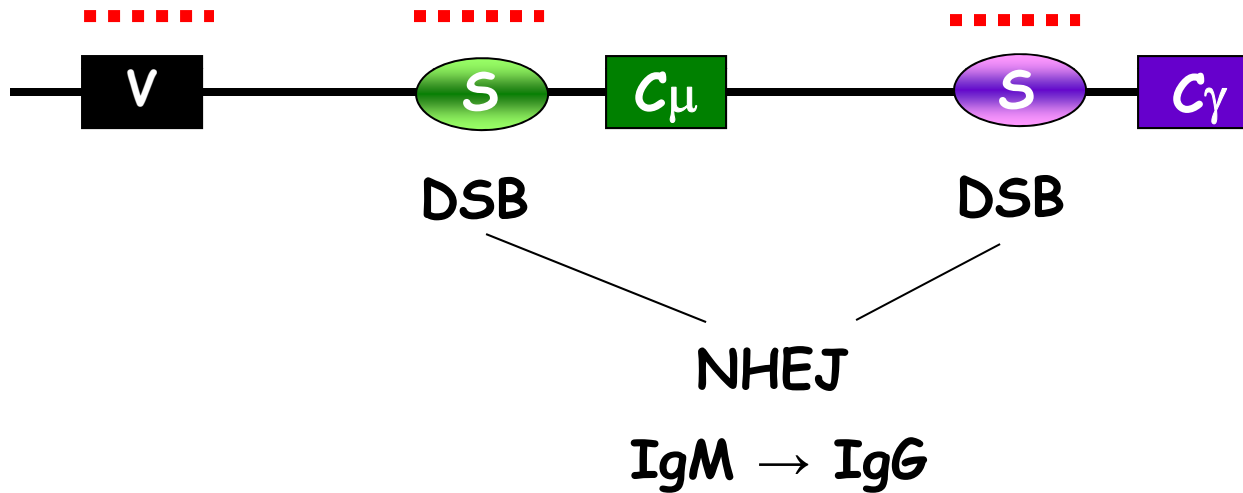
DNA structure pauses RNA pol II and affects mutation

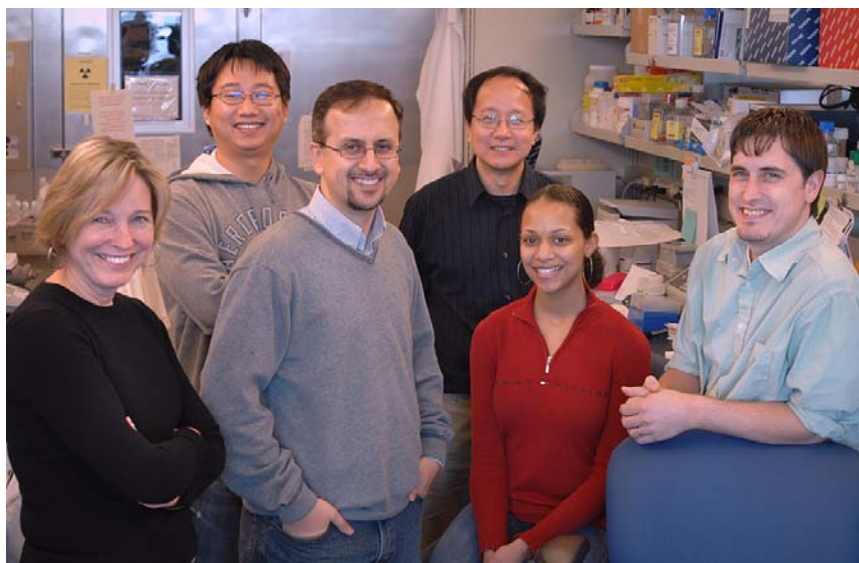
RNA pol II
AID



AID may associate with RNA pol II. R-loop regions pause RNA polymerases, increasing AID binding to ssDNA and mutations. After core, AID falls off or is diluted out, and mutation stops.

What targets AID to V region?





Rob Maul
Huseyin Saribasak
Rhonda McClure
Zheng Cao
William Yang

Rahul Kohli
Jim Stivers
David Wilson
Ranjan Sen
Roger Woodgate