

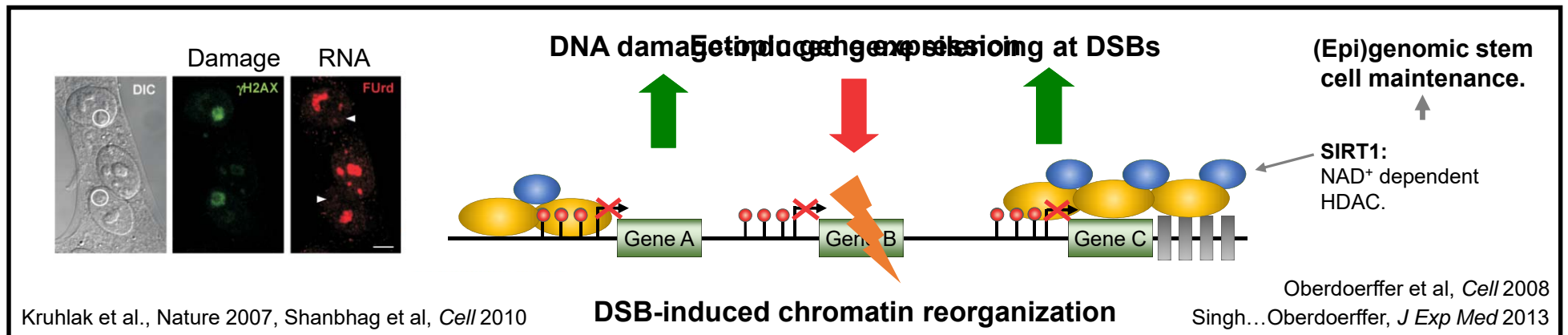
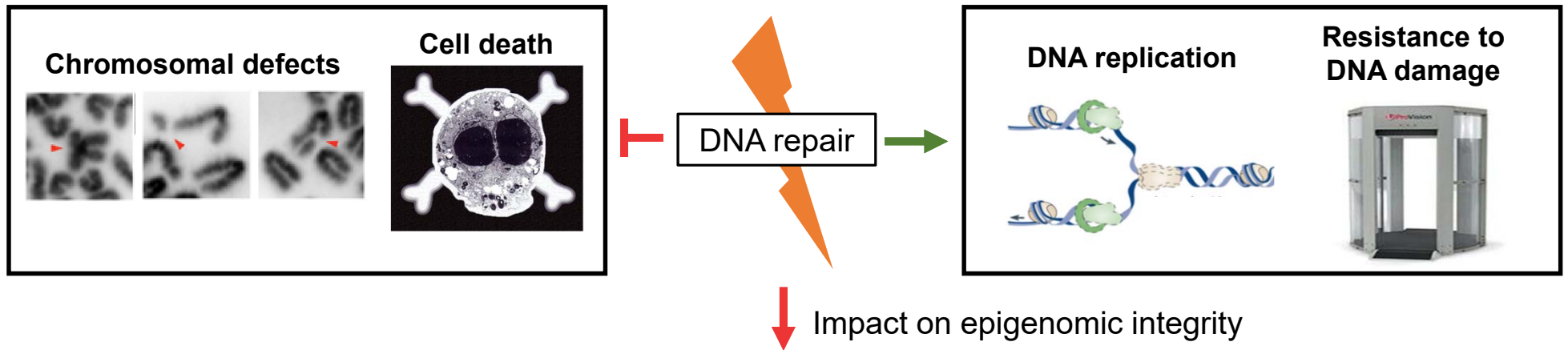


DNA  
damage  
within  
chromatin:  
*break it to  
shape it*

Philipp Oberdoerffer  
NCI, Bethesda, MD

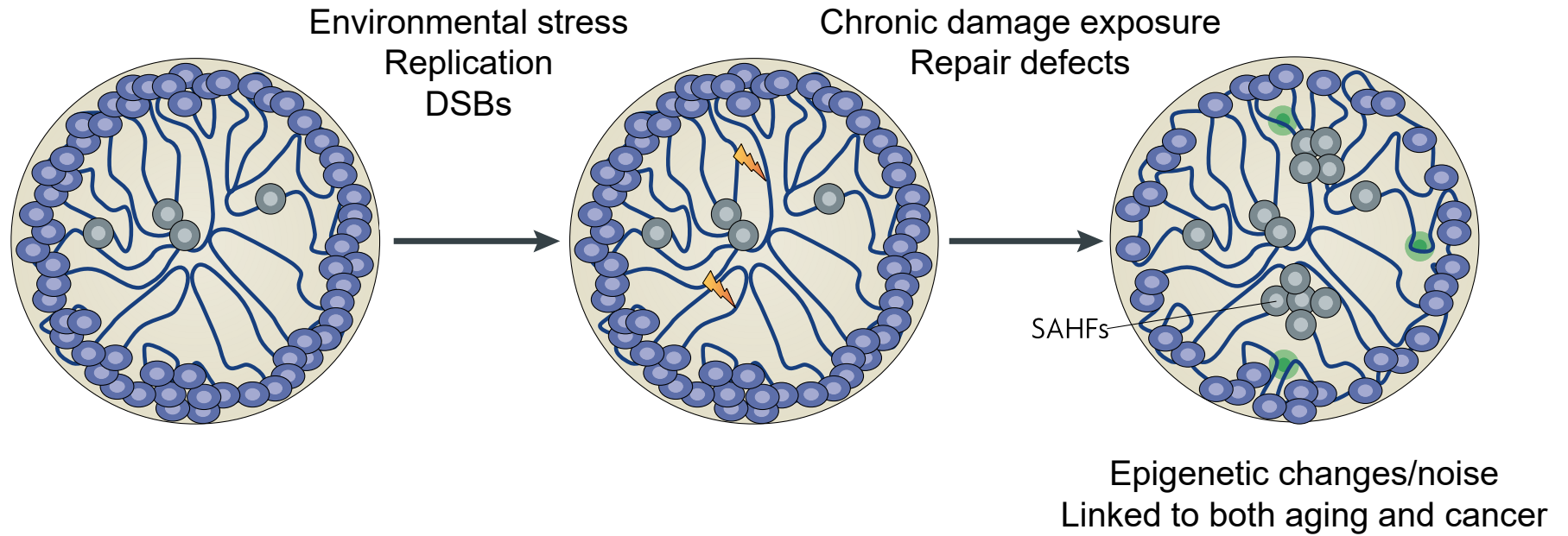
DNA Repair Interest  
Group – 2017

# DNA repair: guarding genome and epigenome integrity?





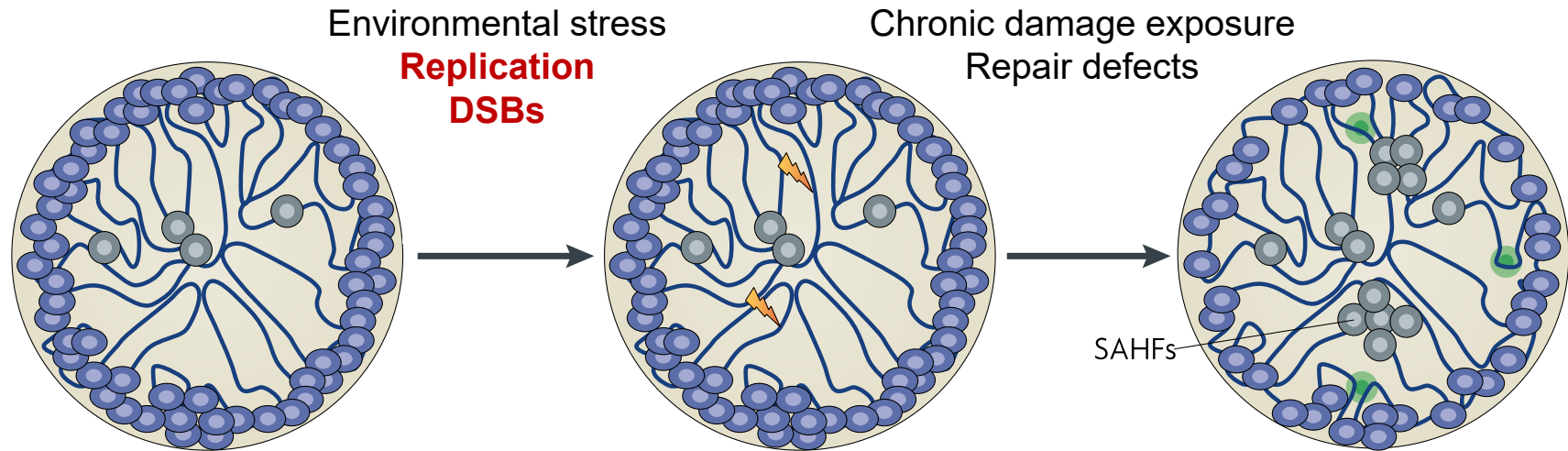
# DNA damage as a driver of epigenomic change in aging and cancer?



Oberdoerffer and Sinclair, "Reorganization of Chromatin Modifiers Hypothesis" *Nature Reviews* 2007

# DNA damage as a driver of epigenomic change in aging and cancer?

---



→ ***Consequences of DSBs for transcriptional integrity in vivo?***

→ ***Chronic DNA damage response as a means to shape chromatin?***

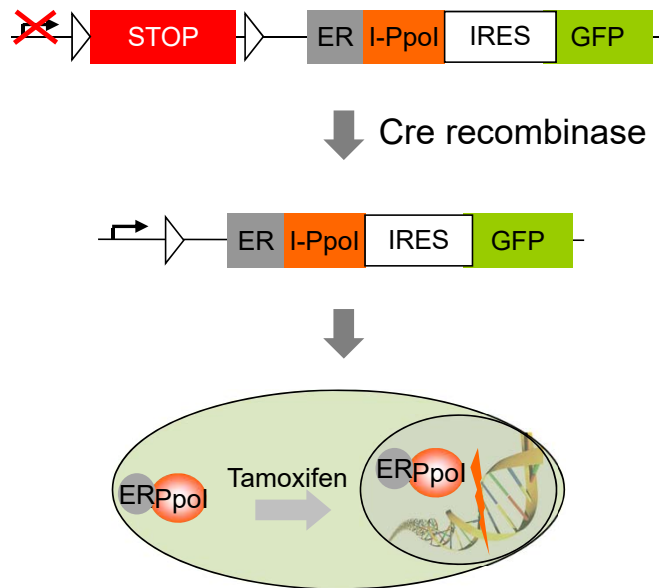
# A mouse model for temporally and spatially controlled DSB induction

## I-Ppol:

Intron-encoded endonuclease from *Physarum polycephalum*

→ Cuts mouse genome at ~ 150 defined sites

## Inducible, tissue-specific I-Ppol mouse model



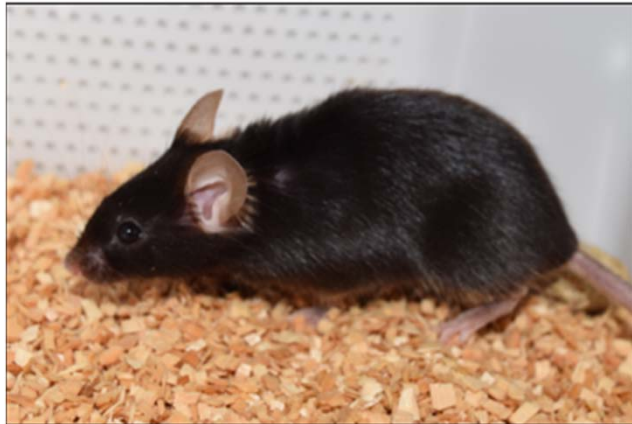
## Transcriptional consequences upon T cell-specific DSB induction?

- I-Ppol causes gene repression specifically in DSB-containing genes.
- DSB-induced gene repression is transient and dependent on DNA damage signaling.

**The mammalian transcriptome is sufficiently robust to accommodate short term DSB exposure.**

## However...

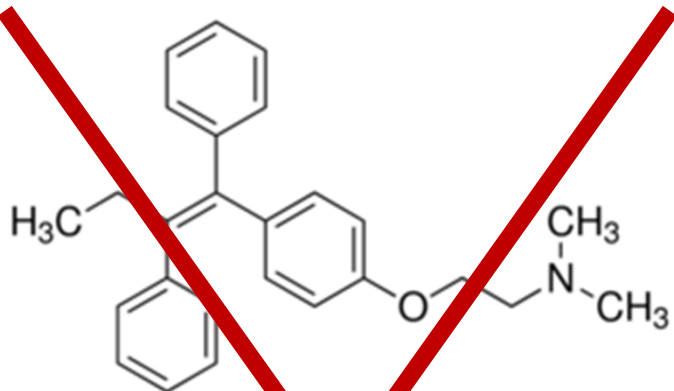
Cre control (16 mo old)



I-Ppo/Cre (16 mo old)



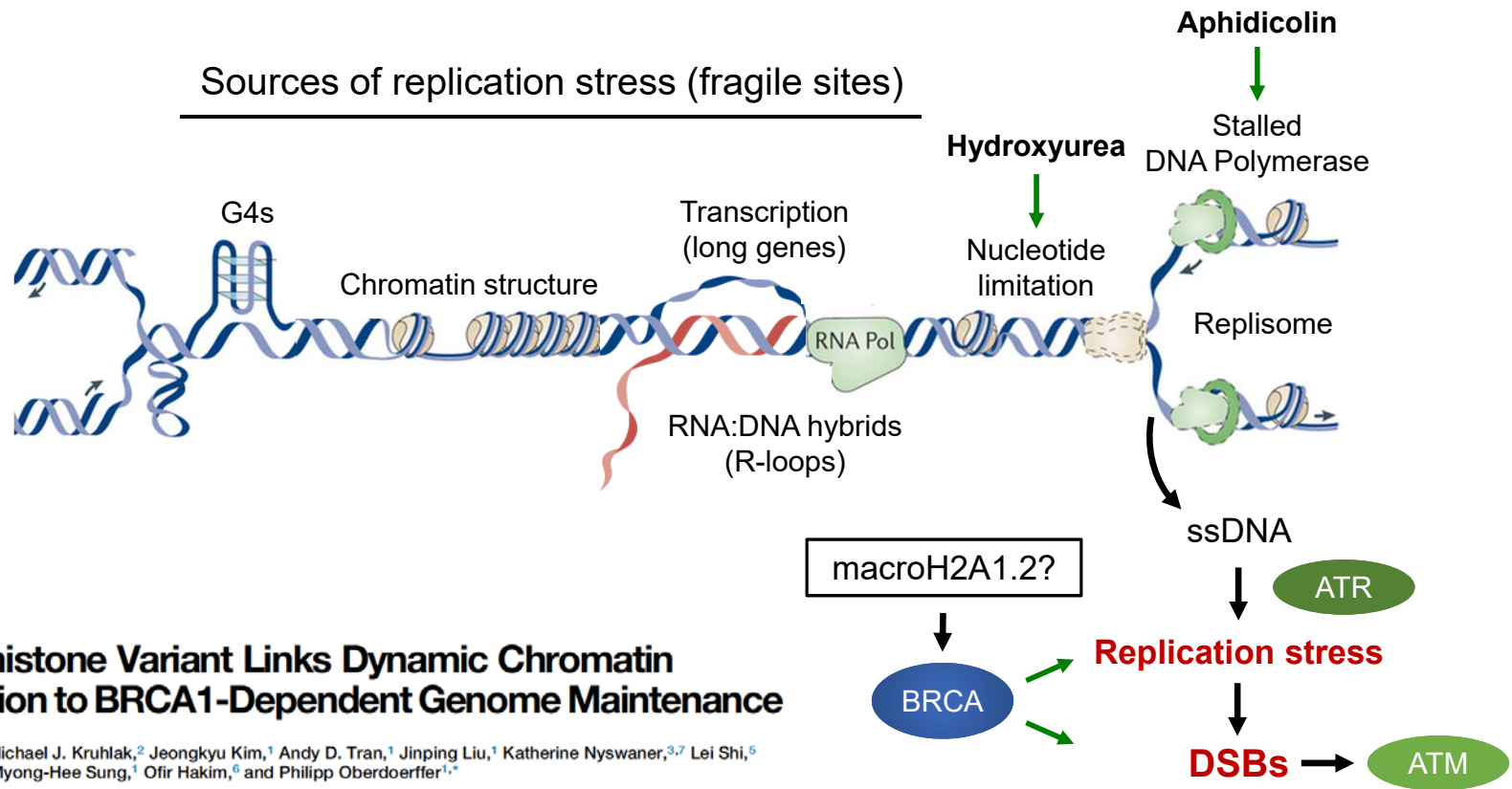
- Mice exposed to several weeks of DSB induction early in life
- Evidence for epigenetic memory of DSB induction...



**Can we test the impact of chronic DNA damage under physiological conditions?**



# Replication stress: a source of chronic, HR-associated DNA damage



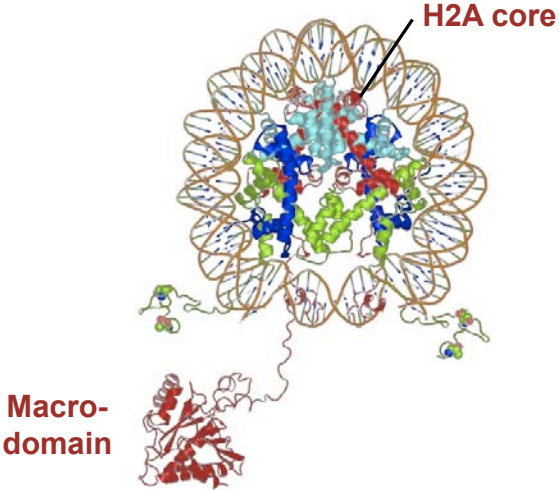
Cell Reports  
Article

## A Macrohistone Variant Links Dynamic Chromatin Compaction to BRCA1-Dependent Genome Maintenance

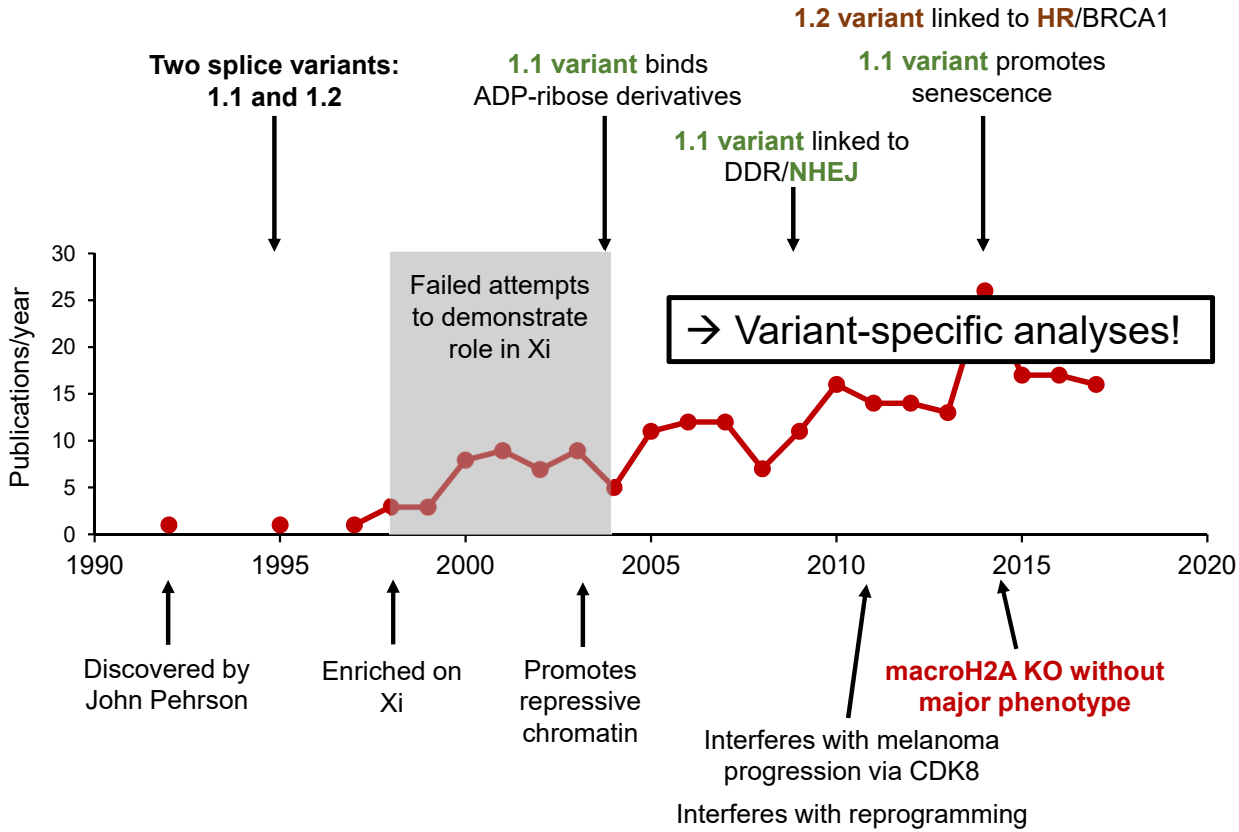
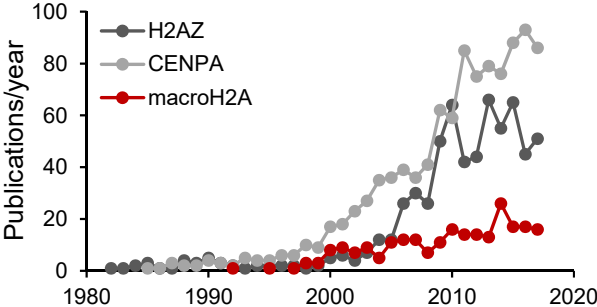
Simran Khurana,<sup>1</sup> Michael J. Kruhlak,<sup>2</sup> Jeongkyu Kim,<sup>1</sup> Andy D. Tran,<sup>1</sup> Jinping Liu,<sup>1</sup> Katherine Nyswaner,<sup>3,7</sup> Lei Shi,<sup>6</sup> Parthav Jailwala,<sup>4</sup> Myong-Hee Sung,<sup>1</sup> Ofir Hakim,<sup>6</sup> and Philipp Oberdoerffer<sup>1,\*</sup>

Adapted from Gaillard...Aguilera, *Nat Rev Cancer* 2015

# MacroH2A1 – a unique yet underappreciated histone variant

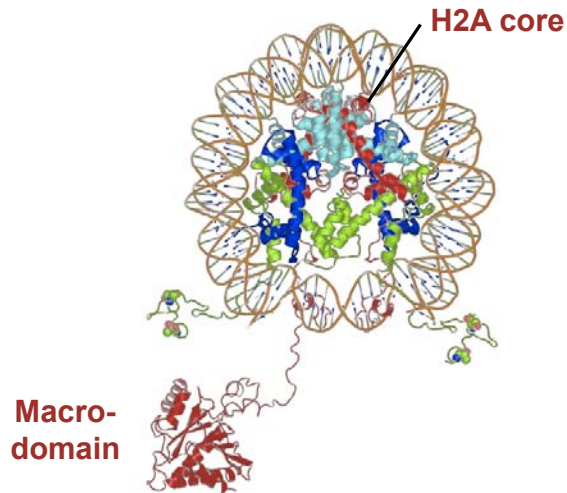


Papers on histone variants



Pehrson, Luger, Bernstein, Ladurner, Gamble, Price, Buschbeck and others...

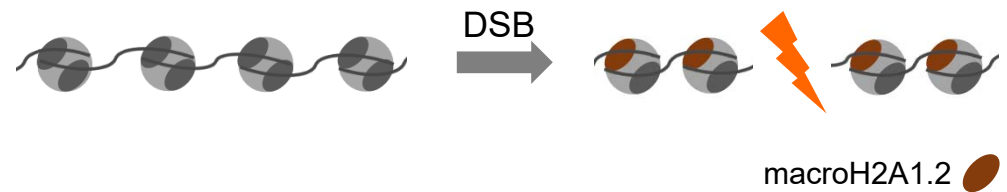
# The macroH2A1.2 variant promotes BRCA1-dependent DSB repair



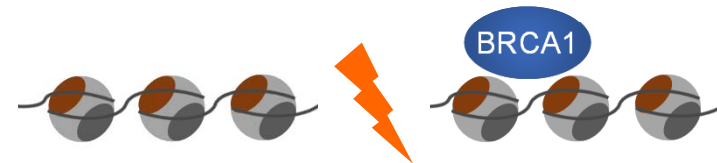
- RNAi screen identified **macroH2A1.2-specific** role in homology-directed DSB repair

Implications for (epi)genome maintenance?

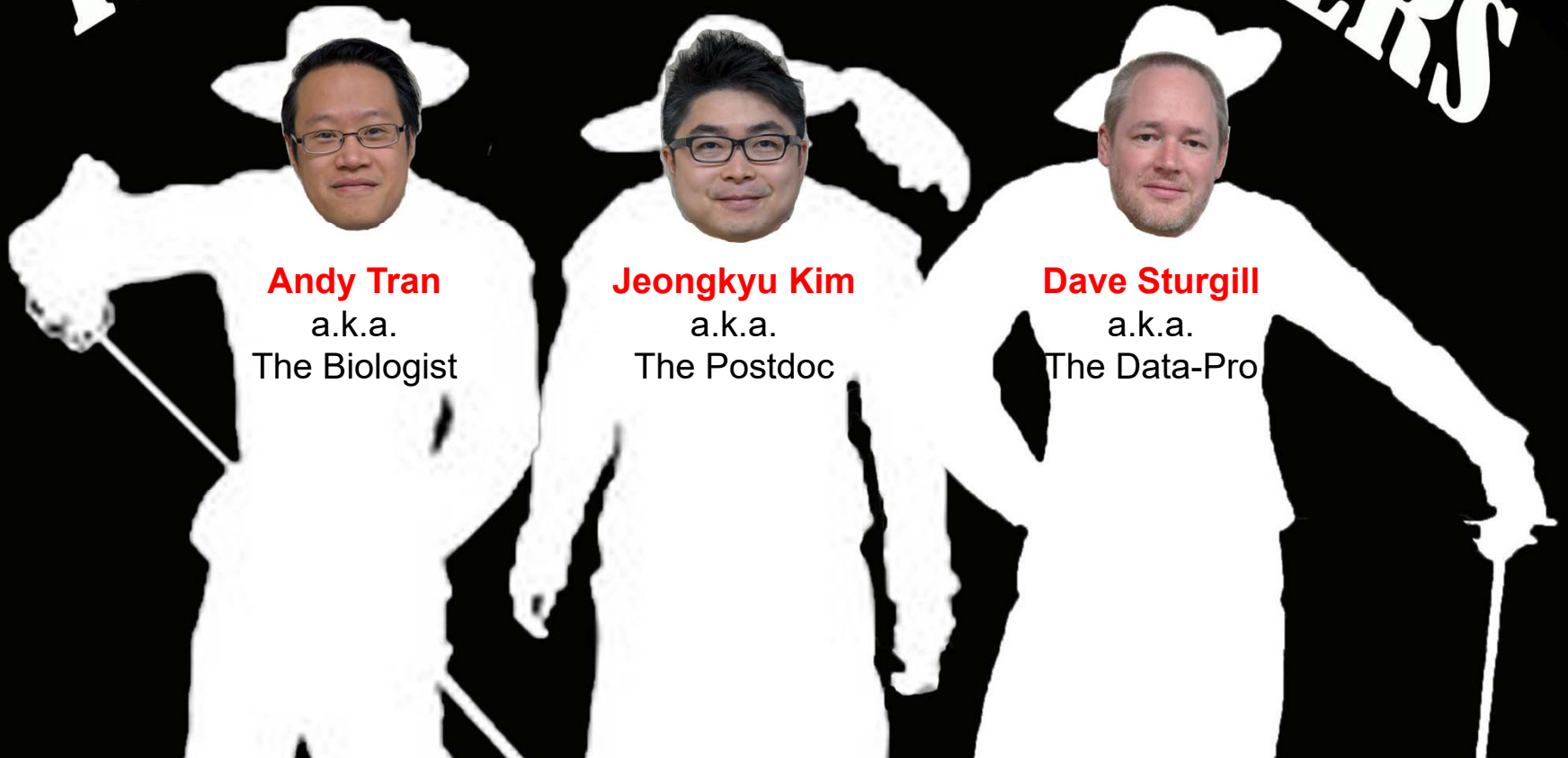
→ **MacroH2A1.2 promotes dynamic chromatin condensation at DSBs**



→ **Facilitates BRCA1 accumulation, end resection and HR**



# THE THREE MUSKETEERS



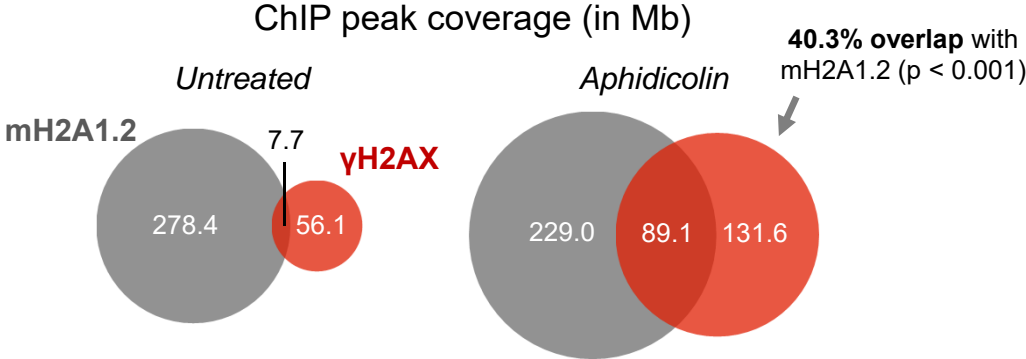
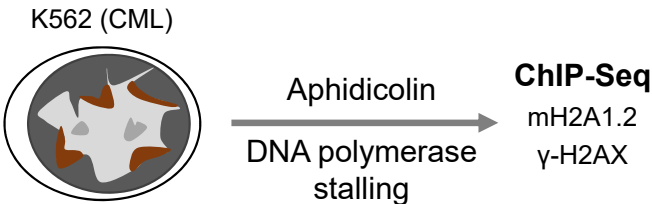
**Andy Tran**  
a.k.a.  
The Biologist

**Jeongkyu Kim**  
a.k.a.  
The Postdoc

**Dave Sturgill**  
a.k.a.  
The Data-Pro

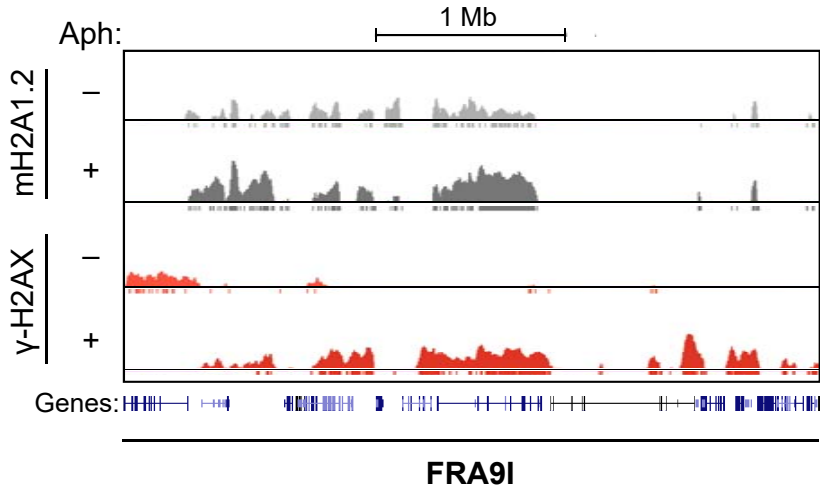
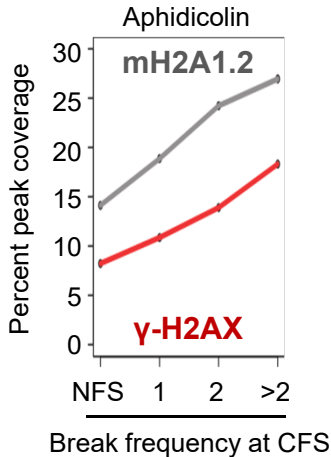
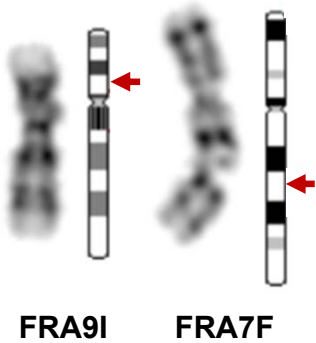
# Replication stress drives macroH2A1.2 accumulation at fragile sites genome-wide

## A role for macroH2A1.2 in replication stress?



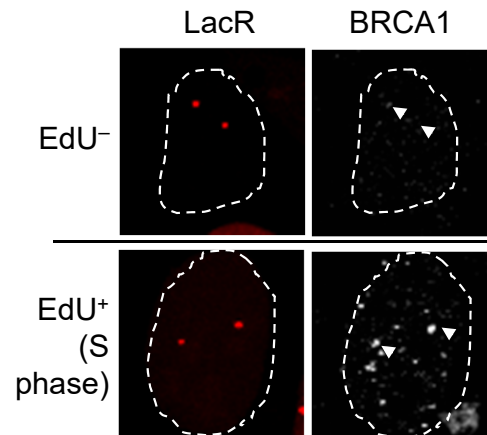
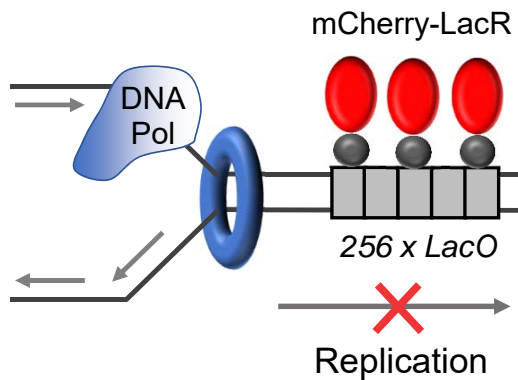
## Relationship to sites of replication stress?

Common fragile sites (CFS)

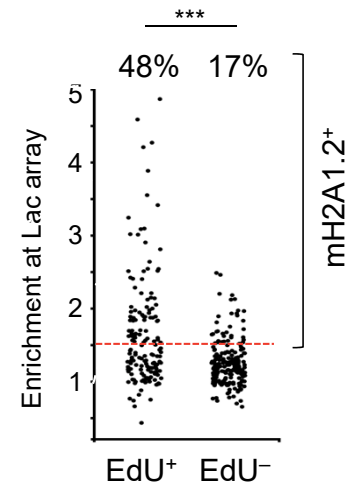
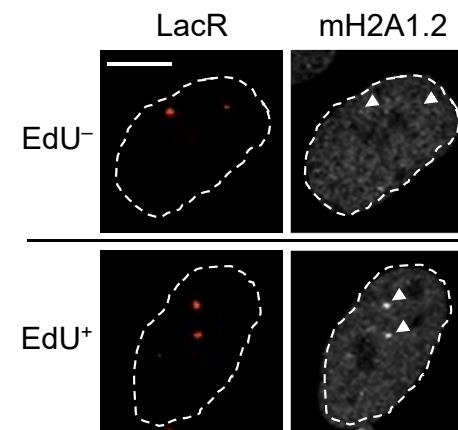


# macroH2A1.2 accumulates at artificial replication blocks

## A system for locally defined replication fork arrest



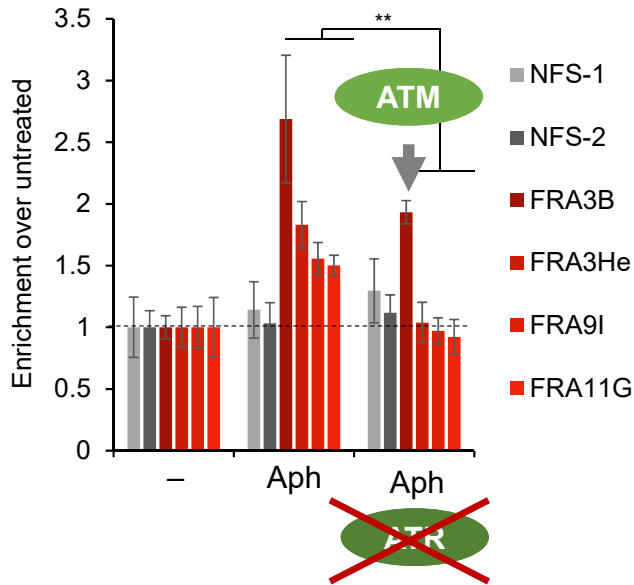
## macroH2A1.2 accumulates at artificial replication stops



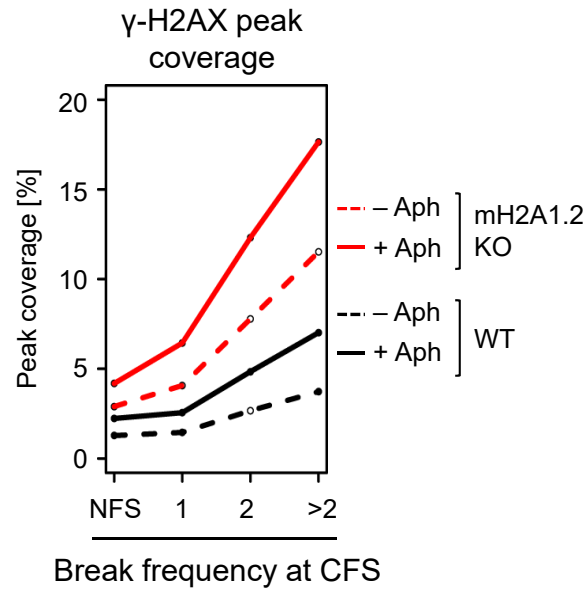
→ Link to the replication stress response?

# Role of macroH2A1.2 during replication stress

## RS-induced mH2A1.2 accumulation is DDR-dependent



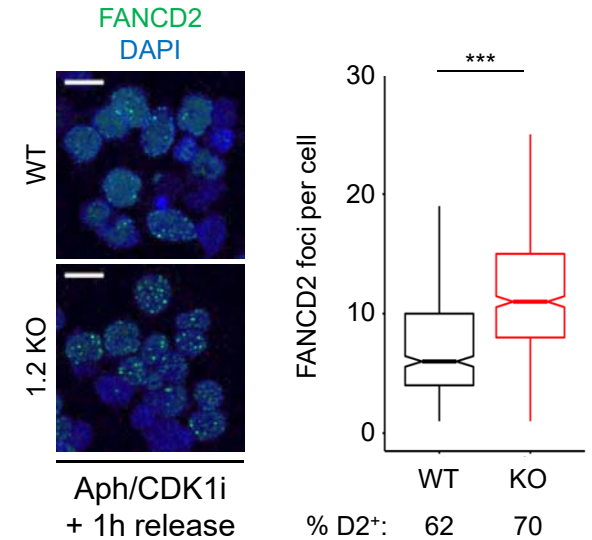
## MacroH2A1.2 loss causes increased DNA damage at CFSs



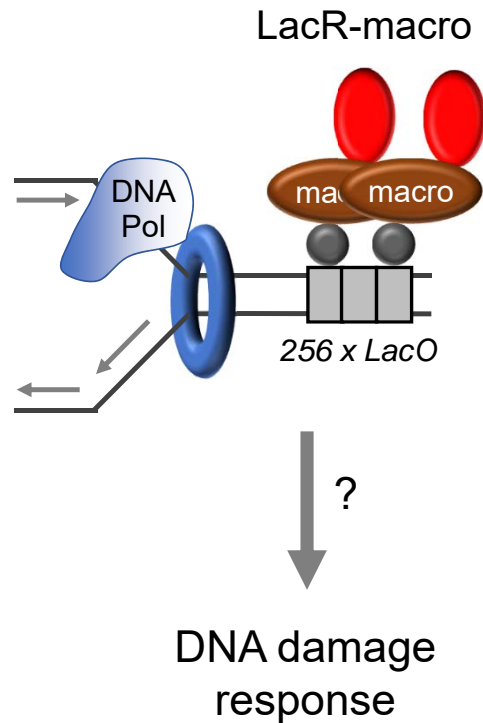
Also by comet assay...

## FANCD2 Facilitates Replication through Common Fragile Sites

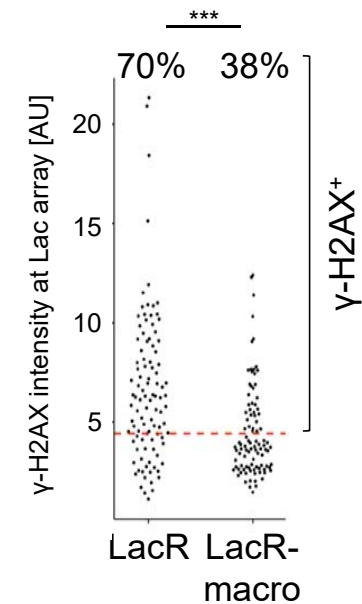
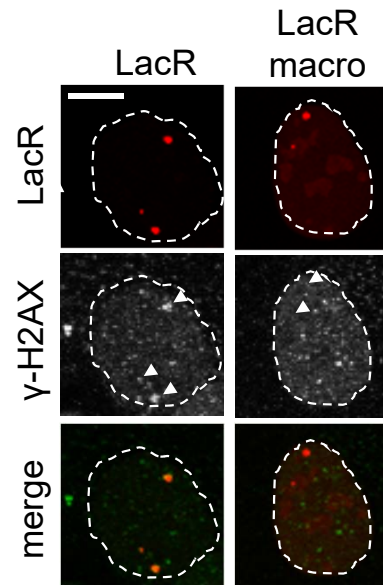
Advaitha Madireddy,<sup>1,\*</sup> Settapong T. Kosiyatrakul,<sup>1</sup> Rebecca A. Boisvert,<sup>2</sup> Emilia Herrera-Moyano,<sup>3</sup> María L. García-Rubio,<sup>3</sup> Jeannine Gerhardt,<sup>1</sup> Elizabeth A. Vuono,<sup>2</sup> Nichole Owen,<sup>2</sup> Zi Yan,<sup>1</sup> Susan Olson,<sup>4</sup> Andrés Aguilera,<sup>3</sup> Niall G. Howlett,<sup>2</sup> and Carl L. Schildkraut<sup>1,5,\*</sup>



# Can macroH2A1.2 protect from replication stress-induced damage?



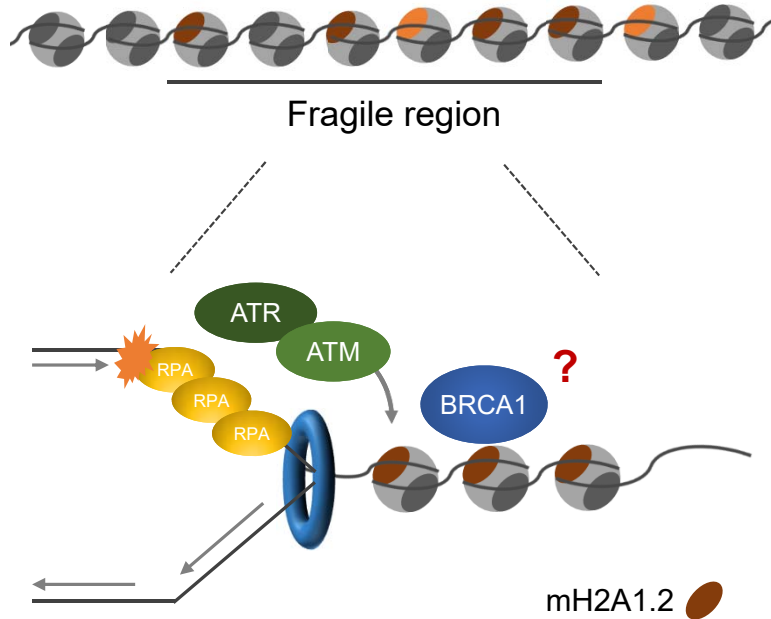
## The macro domain protects from DNA damage at replication blocks



→ Molecular basis for macroH2A1.2-dependent protection from RS?

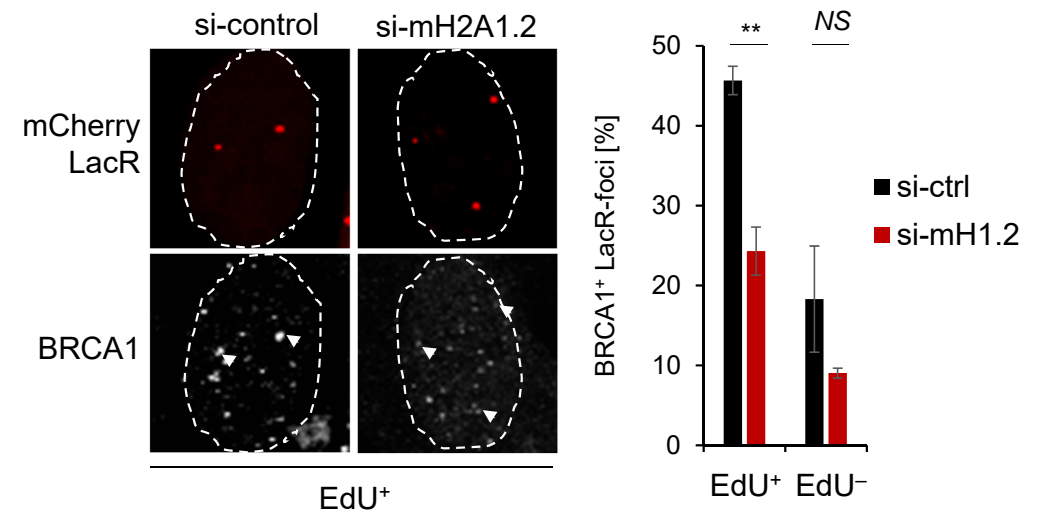


# macroH2A1.2 facilitates BRCA1 accumulation upon replication stress

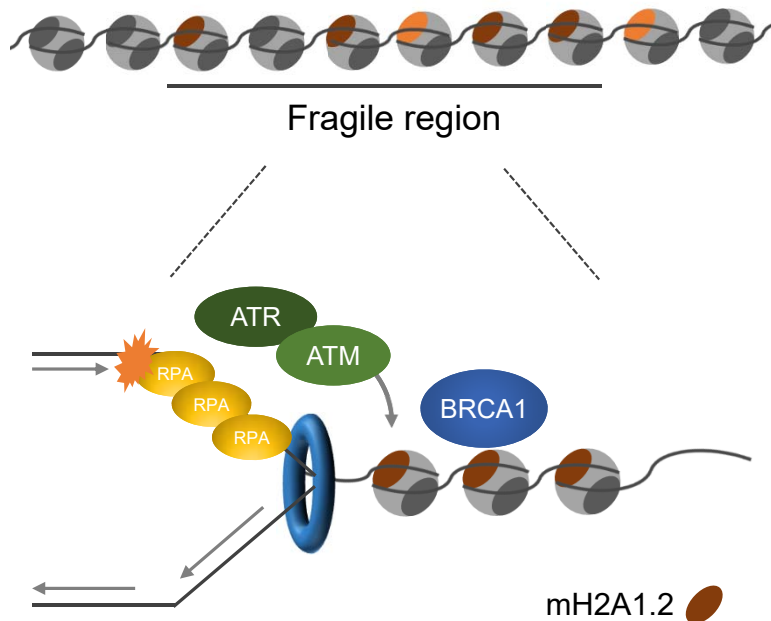


→ Fork specific effect?

## macroH2A1.2 promotes BRCA1 accumulation at replication blocks

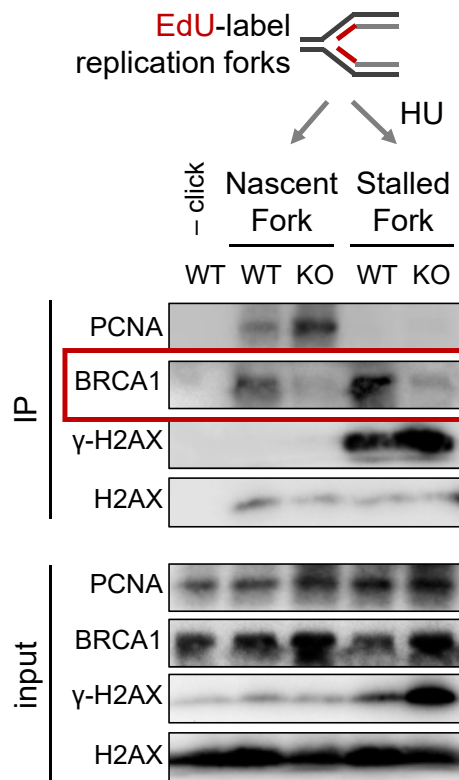


# macroH2A1.2 promotes BRCA1 recruitment to replication forks

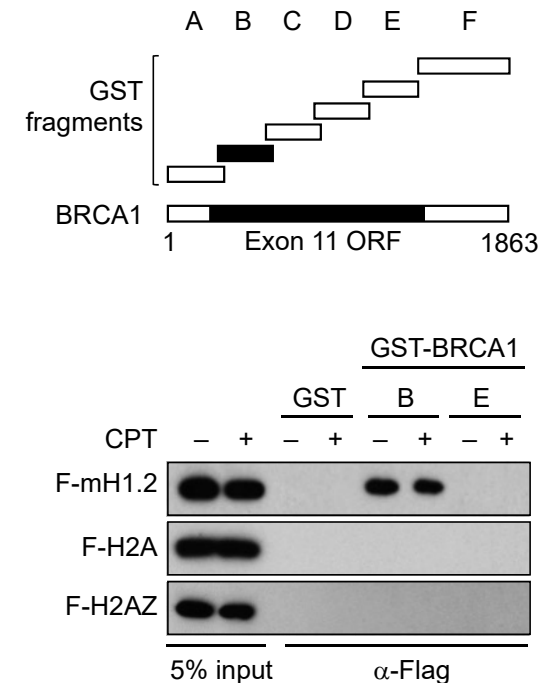


→ What mediates macroH2A1.2 accumulation upon replication stress?

## Isolation of proteins on nascent DNA (iPOND)



## BRCA1 interacts with macroH2A1.2

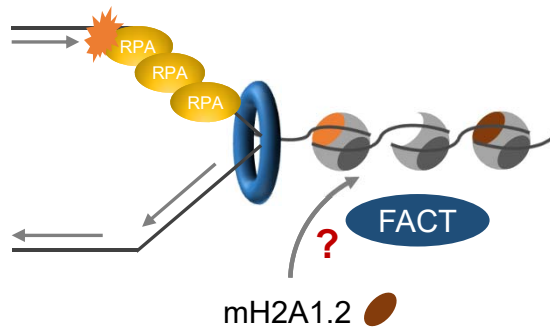


## No known macroH2A1.2 chaperones!

Histone or variant	Tissues (location within the genome)	Chaperones and remodellers	Function
MacroH2A1.1	All (facultative and constitutive heterochromatin)	Not known	Not known
MacroH2A1.2	All (facultative and constitutive heterochromatin)	ATRX (remodeller)	Negative regulation, possibly indirect
MacroH2A2	All (facultative and constitutive heterochromatin)	Not known	Not known

Buschbeck & Hake, Nat. Reviews, 2017

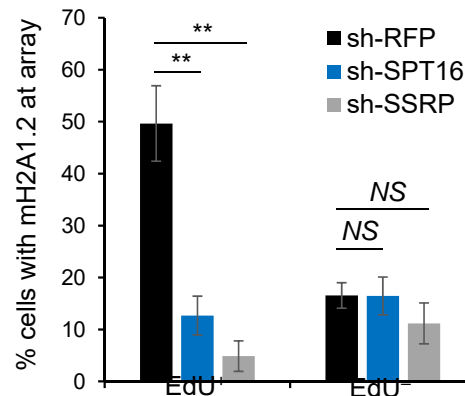
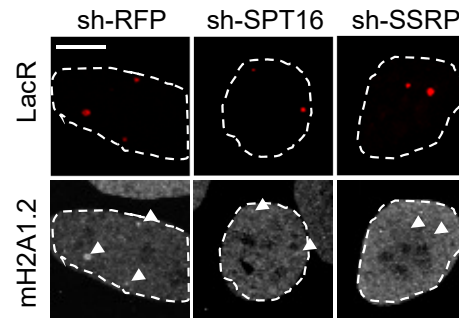
# FACT remodels H2A/H2B dimers at replication forks



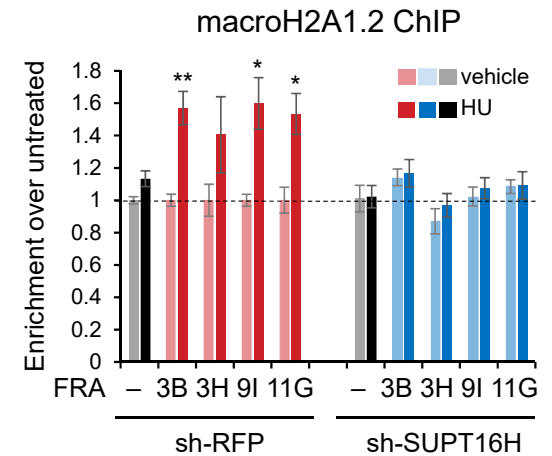
- FACT (SUPT16H/SSRP1) complex is the main H2A/H2B chaperone at replication forks.
- FACT loss causes HR and replication defects.

→ **FACT mediates macroH2A1.2 accumulation in a replication stress-dependent manner.**

FACT subunits promote RS-associated mH2A1.2 accumulation at Lac arrays...



... and common fragile sites.



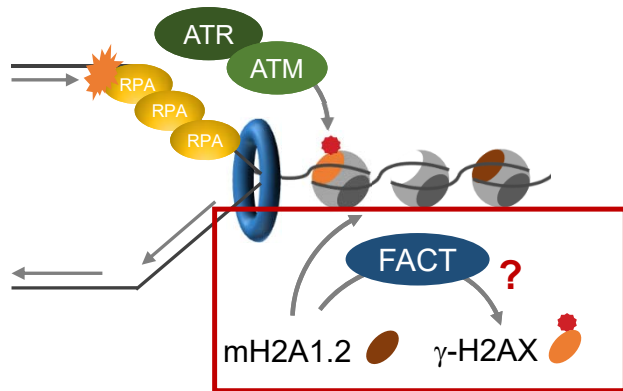
→ FACT interacts with macroH2A1.2/H2B dimer in vitro and in vivo.

# MacroH2A1.2 accumulation at fragile sites requires H2AX phosphorylation

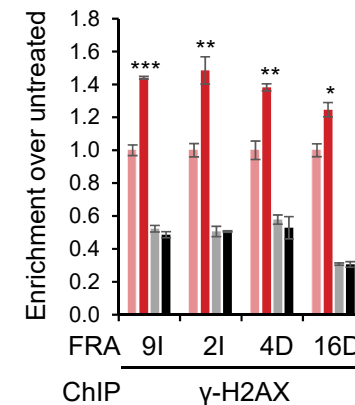
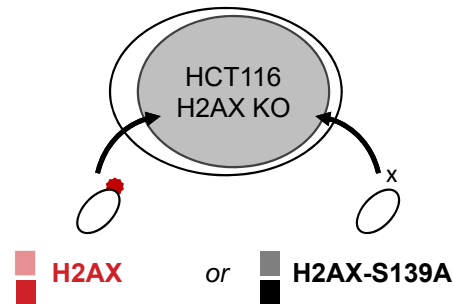
Molecular Cell  
Article

## FACT-Mediated Exchange of Histone Variant H2AX Regulated by Phosphorylation of H2AX and ADP-Ribosylation of Spt16

Kyu Heo,<sup>1</sup> Hyunjung Kim,<sup>1</sup> Si Ho Choi,<sup>2</sup> Jongkyu Choi,<sup>1</sup> Kyunghwan Kim,<sup>1</sup> Jiafeng Gu,<sup>1,3,4,5</sup> Michael R. Lieber,<sup>1,3,4,5</sup> Allen S. Yang,<sup>2</sup> and Woojin An<sup>1,\*</sup>



- FACT (SUPT16H/SSRP1) complex is the main H2A/H2B chaperone at replication forks.
  - FACT loss causes HR and replication defects.
- **FACT mediates macroH2A1.2 accumulation in a replication stress-dependent manner.**
- **MacroH2A1.2 incorporation also depends on DNA damage signaling.**



→ MacroH2A1.2 incorporation requires H2AX phosphorylation.

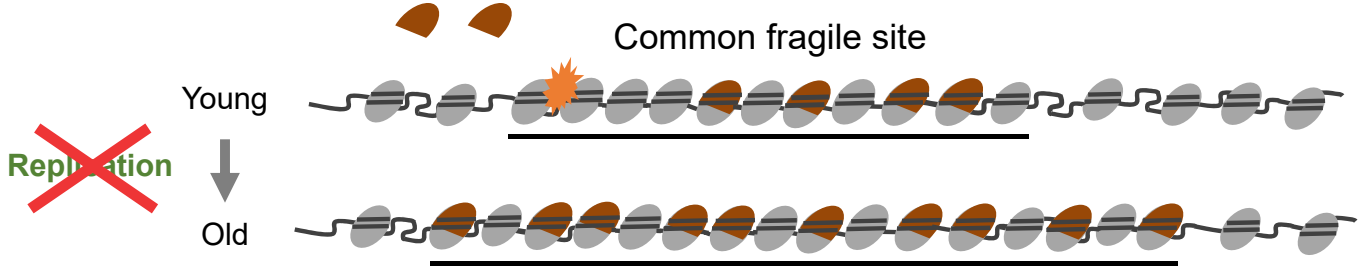
## Potential to shape chromatin during continued replication stress?

---



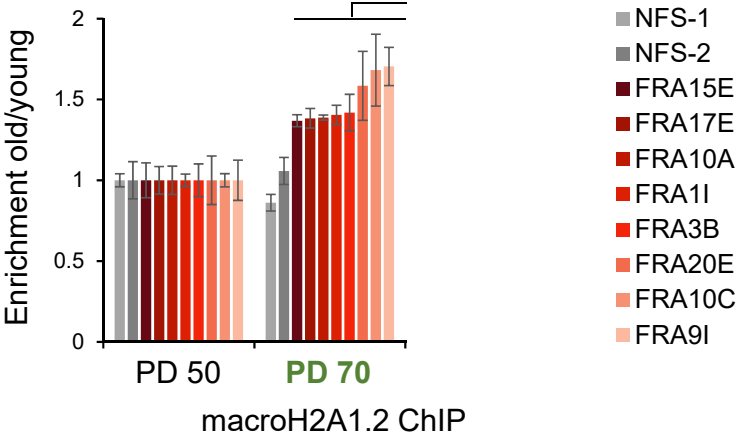
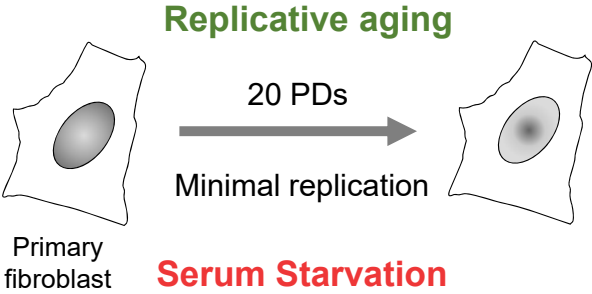
The Lego problem...

# Continuous replication drives persistent macroH2A1.2 accumulation

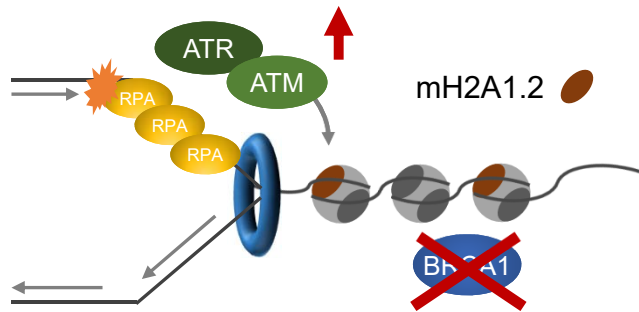


→ Dependence on replication? ✓

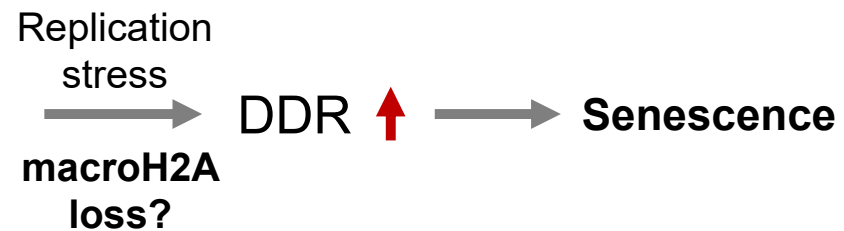
Replicative age causes mH2A1.2 accumulation at CFSs



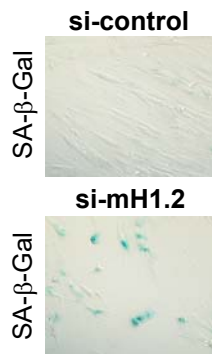
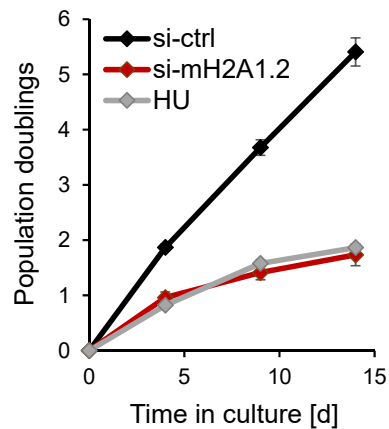
# Consequences of macroH2A1.2 loss in primary cells



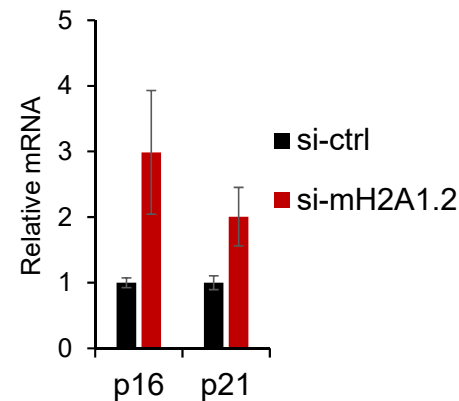
→ **Barrier to malignant transformation**



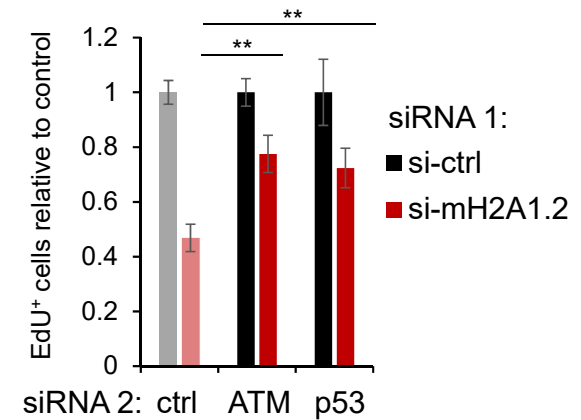
macroH2A loss promotes senescence



DDR is activated



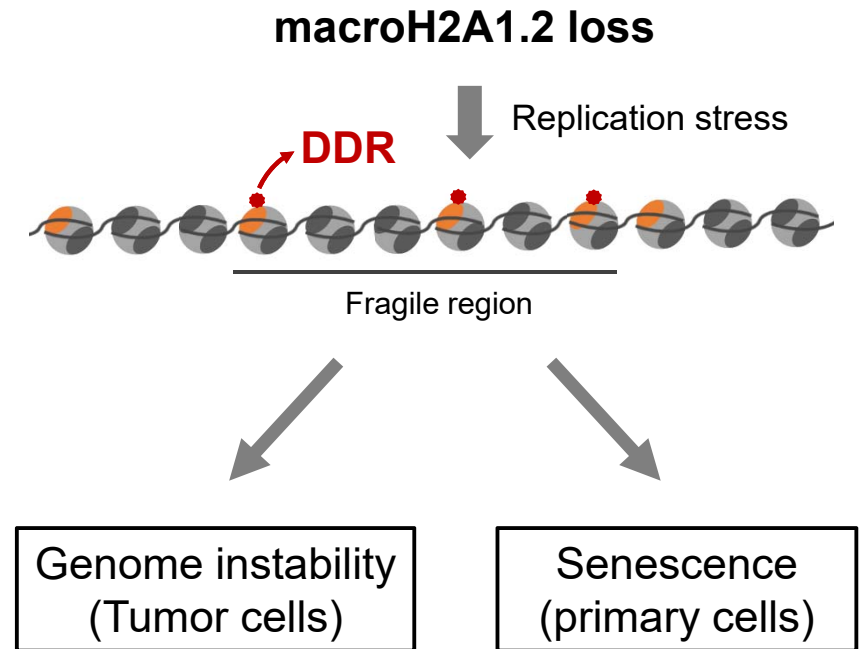
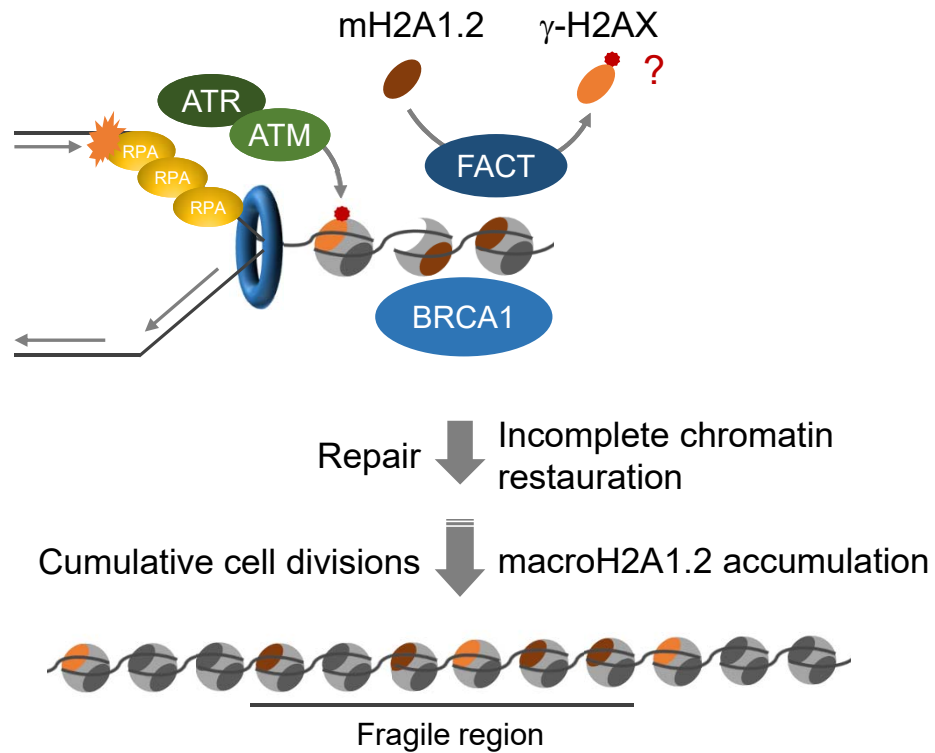
Cell cycle arrest is DDR-dependent



Bartek, d'Adda di Fagagna and others

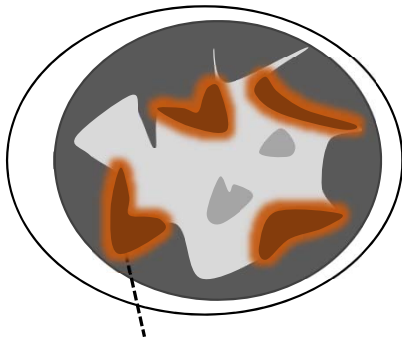


# Replication stress shapes a protective chromatin environment at fragile regions



## Implications for the epigenome

---

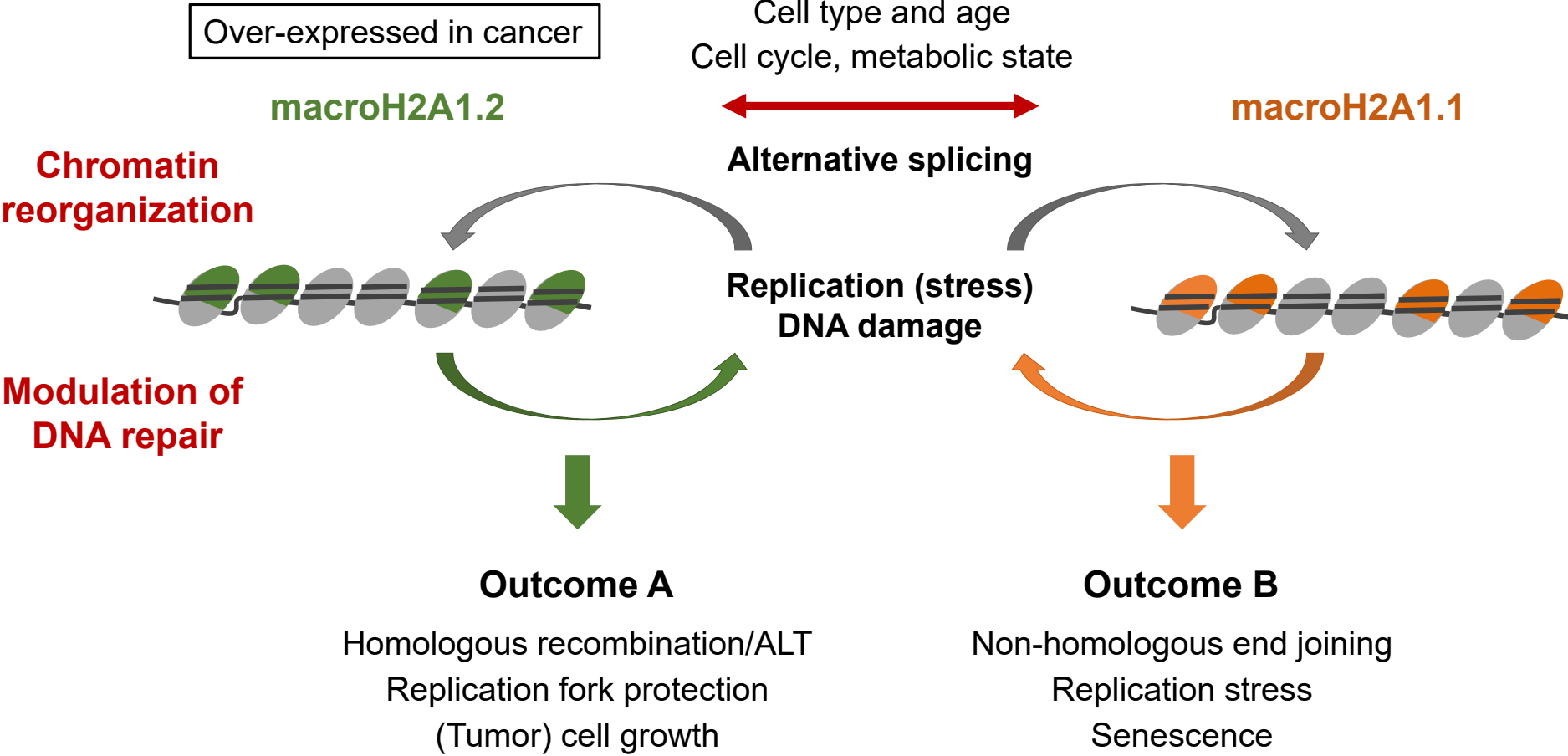


Fragile regions / macroH2A1.2 domains

- **A subset of chromatin domains are the result of chronic DNA damage exposure.**
- **Implications for age-associated chromatin reorganization and epigenomic dysfunction.**

**A more general role for macroH2A1.2 in HR and/or replication stress-associated genome maintenance?**

# Feedback between DNA repair and chromatin modulates cell function



# Acknowledgements



## Lab members:

Jeongkyu Kim  
Robin Sebastian  
Eri Hosogane  
Will Hannon  
Alex Kruswick  
David Sturgill

## Collaborators:

*Bill Bonner (NCI)*  
*Urbain Weyemi*

*Sandy Burkett (NCI)*

*Karolin Luger (UC Boulder)*  
*Garrett Edwards*

*Yie Liu (NIA)*  
*Chongkiu Sun*  
*Pei-Ji Chin*



## Former:

Simran Khurana  
Jinping Liu  
Alex Kruswick



**National Cancer Institute**  
Center for Cancer Research